

UNIVERSIDADE FEDERAL DE GOIÁS
FACULDADE DE NUTRIÇÃO
PROGRAMA DE PÓS-GRADUAÇÃO EM NUTRIÇÃO E SAÚDE

AMANDA GOULART DE OLIVEIRA SOUSA

**QUALIDADE NUTRICIONAL E VALOR PROTÉICO DAS
AMÊNDOAS DE BARU, DE PEQUI E DA CASTANHA-DE-
CAJU-DO-CERRADO EM RELAÇÃO AO AMENDOIM**

Goiânia
2011

**Dados Internacionais de Catalogação na Publicação na (CIP)
GPT/BC/UFG**

S725q Sousa, Amanda Goulart.
Qualidade nutricional e valor nutritivo das amêndoas de baru,
de pequi e da castanha-de-caju-do-cerrado em relação ao
amendoim [manuscrito] / Amanda Goulart de Oliveira Sousa. -
2011.
54 f. : il., figs, tabs.

Orientadora: Prof^a. Dr^a. Maria Margareth Veloso Naves.
Dissertação (Mestrado) – Universidade Federal de Goiás,
Faculdade de Nutrição, 2011.
Bibliografia.
Anexos.

1. Qualidade nutricional. 2. Valor protéico. 3. Amêndoas. 4.
Noz. I. Título.

CDU: 634.55:634.58

AMANDA GOULART DE OLIVEIRA SOUSA

**QUALIDADE NUTRICIONAL E VALOR PROTÉICO DAS
AMÊNDOAS DE BARU, DE PEQUI E DA CASTANHA-DE-
CAJU-DO-CERRADO EM RELAÇÃO AO AMENDOIM***

Dissertação de Mestrado apresentada ao Programa de Pós-Graduação em Nutrição e Saúde da Faculdade de Nutrição da Universidade Federal de Goiás para obtenção do Título de Mestre em Nutrição e Saúde.

Orientadora: Prof^a Dr^a Maria Margareth Veloso Naves

Linha de pesquisa: Qualidade de alimentos e dietas

Goiânia
2011

*Este trabalho é parte do projeto financiado pelo Conselho Nacional de Desenvolvimento Científico e Tecnológico (CNPq/ Edital Universal CNPq - 2008).

TERMO DE CIÊNCIA E DE AUTORIZAÇÃO PARA DISPONIBILIZAR AS TESES E DISSERTAÇÕES ELETRÔNICAS (TEDE) NA BIBLIOTECA DIGITAL DA UFG

Na qualidade de titular dos direitos de autor, autorizo a Universidade Federal de Goiás (UFG) a disponibilizar, gratuitamente, por meio da Biblioteca Digital de Teses e Dissertações (BDTD/UFG), sem ressarcimento dos direitos autorais, de acordo com a Lei nº 9610/98, o documento conforme permissões assinaladas abaixo, para fins de leitura, impressão e/ou download, a título de divulgação da produção científica brasileira, a partir desta data.

1. Identificação do material bibliográfico: ☒ **Dissertação** ☐ **Tese**

2. Identificação da Tese ou Dissertação

Autor (a):	Amanda Goulart de Oliveira Sousa		
E-mail:	goulart.amanda@gmail.com		
Seu e-mail pode ser disponibilizado na página? ☒ Sim ☐ Não			
Vínculo empregatício do autor	Pontifícia Universidade Católica de Goiás		
Agência de fomento:		Sigla:	PUC-GO
País:	Brasil	UF:GO	CNPJ: 01.587.609/0001-71
Título:	Qualidade nutricional e valor protéico das amêndoas de baru, de pequi e da castanha-de-caju-do-cerrado em relação ao amendoim		
Palavras-chave:	Frutos nativos, Cerrado, nozes, amêndoas, composição química, qualidade protéica		
Título em outra língua:	Nutritional quality and protein value of exotic almonds and nut from the Brazilian Savanna compared to peanut		
Palavras-chave em outra língua:	Native fruits, Brazilian Savanna, nut, almond, chemical composition, protein quality		
Área de concentração:	Nutrição		
Data defesa:	01/04/2011		
Programa de Pós-Graduação:	Nutrição e Saúde		
Orientador (a):	Dra. Maria Margareth Veloso Nunes		
E-mail:	mnunes@fanut.ufg.br		

3. Informações de acesso ao documento:

Liberção para disponibilizaço?¹ ☒ total ☐ parcial

Em caso de disponibilizaço parcial, assinale as permissões:

☐ Capítulos. Especifique:

☐ Outras restrições: _____

Havendo concordância com a disponibilizaço eletrônica, torna-se imprescindível o envio do(s) arquivo(s) em formato digital PDF ou DOC da tese ou dissertaço.

O Sistema da Biblioteca Digital de Teses e Dissertaço garante aos autores, que os arquivos contendo eletronicamente as teses e ou dissertaço, antes de sua disponibilizaço, receberão procedimentos de segurança, criptografia (para não permitir cópia e extraço de conteúdo, permitindo apenas impresso fraca) usando o padrão do Acrobat.

Assinatura do (a) autor (a)

Data: ____ / ____ / ____

¹ Em caso de restrição, esta poderá ser mantida por até um ano a partir da data de defesa. A extensão deste prazo suscita justificativa junto à coordenação do curso. Todo resumo e metadados ficarão sempre disponibilizados.

UNIVERSIDADE FEDERAL DE GOIÁS
FACULDADE DE NUTRIÇÃO
PROGRAMA DE PÓS-GRADUAÇÃO EM NUTRIÇÃO E SAÚDE

AMANDA GOULART DE OLIVEIRA SOUSA

**QUALIDADE NUTRICIONAL E VALOR PROTÉICO DAS
AMÊNDOAS DE BARU, DE PEQUI E DA CASTANHA-DE-
CAJU-DO-CERRADO EM RELAÇÃO AO AMENDOIM**

Dissertação DEFENDIDA e APROVADA em 01 de abril de 2011, pela Banca Examinadora constituída pelos membros:

Profª Drª Maria Célia Lopes Torres
Membro – EA/ UFG

Profª Drª Mara Reis Silva
Membro – FANUT/ UFG

Profª Drª Maria Margareth Veloso Naves
Orientadora

*Dedico este trabalho primeiramente à Deus.
À minha mãe, Waldeci, pelo seu esforço, dedicação e compreensão.
Às minhas irmãs, Sandra e Áurea, meus exemplos.
Ao meu esposo, Paulo, pelo incentivo, paciência e apoio em todos os momentos.
À minha orientadora, Margareth, por me ajudar a realizar este sonho.*

AGRADECIMENTOS

À Deus por ter me concedido a vida, saúde e disposição.

À minha mãe Waldeci, a, que sempre me estimulou a estudar e não mediu esforços para que eu tivesse uma boa educação.

Ao meu esposo, Paulo Gustavo Pedreira e Sousa, que sempre me apoiou e compreendeu.

Às minhas irmãs, Sandra de Fátima Oliveira e Aurea Maria de Oliveira Pitaluga, que sempre foram meus exemplos de formação pessoal e intelectual.

À Prof^a Dr^a Maria Margareth Veloso Naves (Faculdade de Nutrição - UFG), pela orientação na iniciação científica durante a graduação, o que me estimulou a trilhar esse caminho e também pela sua excelente orientação que me prestou durante esta nova etapa.

À amiga de mestrado e Nutricionista Daniela Canuto Fernandes, pelo auxílio durante todos os momentos do desenvolvimento da pesquisa, um verdadeiro “anjo” que Deus enviou para me ajudar.

Ao técnico Tiago Dias (LANAAL – FANUT/UFG) pelo apoio nas análises químicas.

À nutricionista e mestre em Ciência e Tecnologia de Alimentos, Jullyana Borges de Freitas, pelo apoio e contribuições ao desenho e desenvolvimento da pesquisa.

Às minhas colegas de mestrado Eloísa Helena Carrijo Barbosa e Lorena Pereira de Souza Rosa pelo apoio durante o ensaio biológico.

Ao Prof. Dr. Moacir Evandro Lage e ao técnico de laboratório Rodrigo Almeida de Oliveira (Escola de Veterinária - UFG) pelas análises de ácidos graxos.

Ao Técnico Luiz Sávio Teixeira (LANAGRO - GO) pelas análises de minerais.

À Prof^a Dr^a Sílvia Maria Franciscato Cozzolino e ao Técnico de Laboratório José Alexandre Pimentel (Faculdade de Ciências Farmacêuticas - USP) pelas análises de selênio.

À Técnica Clarice Izumi (Centro de Química de Proteínas da Faculdade de Medicina de Ribeirão Preto – USP) pelas análises de aminoácidos.

À ex-aluna do curso de graduação em Nutrição da UFG, Aline Medeiros Alves, pelo apoio na coleta das amostras, nas análises químicas e no ensaio biológico.

Às ex-alunas do curso de graduação em Nutrição da PUC-GO, Bruna Aniele Cota, Camila Kellen de Souza Cardoso e Pamella Tayná Moreira e Silva, pelo auxílio no ensaio biológico.

À aluna do curso de graduação em Nutrição da PUC-GO, Camila Pinto Lemos, pelo auxílio no ensaio biológico.

À Fundação de Amparo à Pesquisa do Estado de Goiás (FAPEG) pela concessão de bolsa de pós-graduação.

Ao Conselho Nacional de Desenvolvimento Científico e Tecnológico (CNPq/ Edital Universal CNPq - 2008) pelo auxílio financeiro ao projeto.

RESUMO

O objetivo deste estudo foi determinar a qualidade nutricional e o valor protéico das amêndoas de baru e de pequi e da castanha-de-caju-do-cerrado, frutas nativas do Cerrado brasileiro, e comparar com o amendoim. Determinou-se a composição química centesimal, teor de minerais e perfil de aminoácidos, conforme métodos padronizados. Foi realizado um experimento com 42 ratos Wistar, machos, recém-desmamados, distribuídos em sete grupos segundo delineamento por blocos casualizados, durante catorze dias. As dietas foram formuladas segundo AIN-93G, sendo seis dietas com 10% de proteína: CAS7 (caseína com 7% de lipídios); CAS15 (caseína com 15% de lipídios); amêndoas de baru (AMB); amêndoa de pequi (AMP); castanha-de-caju-do-cerrado (CJC) e amendoim (AMD), e uma dieta aprotéica (APO). O valor protéico foi estimado por meio dos métodos *Net Protein Ratio* (NPR), *Relative Net Protein Ratio* (RNPR) e *Protein Digestibility-Corrected Amino Acid Score* (PDCAAS). As amêndoas de baru, de pequi e a castanha-de-caju-do-cerrado são ricas em proteínas (22,7-29,9 g/100 g), lipídios (41,9-50,0 g/100 g), fibras (amêndoas de baru e de pequi, em torno de 10,0 g/100 g), ferro e zinco (4,3-7,4 mg/100 g). A proteína da amêndoa de baru não apresentou deficiência em aminoácidos essenciais, e a lisina foi o primeiro aminoácido limitante nas proteínas da amêndoa de pequi e da castanha-de-caju-do-cerrado, e o segundo limitante na proteína do amendoim. A amêndoa de baru apresentou RNPR de 86%, estatisticamente similar ao da castanha-de-caju-do-cerrado (78%), mas superior ao do amendoim (72%) e da amêndoa do pequi (54%). A amêndoa de baru apresentou maior valor de PDCAAS (91%), e a castanha-de-caju-do-cerrado e o amendoim apresentaram valores semelhantes para este índice (82%), seguidos pela amêndoa de pequi (55%). A amêndoa de baru possui maior qualidade protéica, porém a castanha-de-caju-do-cerrado e o amendoim também são fontes de proteína de boa qualidade. Recomendamos a inclusão destes alimentos nativos em dietas saudáveis e na indústria alimentícia, e a amêndoa de baru e a castanha-de-caju-do-cerrado como fontes de proteínas complementares.

Palavras-chave: Frutos nativos, Cerrado, nozes, amêndoas, composição química, qualidade protéica.

ABSTRACT

The aim of this study was to determine the nutritional quality and protein value of the baru almond, pequi almond, and cerrado cashew nut, native fruits from the Brazilian Savanna, compared to the peanut. Standardized methods were used to determine centesimal composition, amino acid profile, fatty acids and mineral content. The experiment was carried out with 42 male weanling Wistar rats. The animals were randomly assigned into seven groups. The experiment lasted fourteen days. The diets were formulated according to AIN-93G, six diets with 10% protein: CAS7 (7% lipid casein), CAS15 (15% lipid casein), AMB (baru almond), AMP (pequi almond), CJC (cerrado cashew nut), AMD (peanut) and a protein-free diet. A biological assay was carried out to assess the protein value, by Net Protein Ratio (NPR), Relative Net Protein Ratio (RNPR), and Protein Digestibility-Corrected Amino Acid Score (PDCAAS) methods. We found that the exotic almonds and the nut are rich in proteins (22.7–29.9 g/100 g), lipids (41.9–50.0 g/100 g), fibres (baru and pequi almonds, around 10.0 g/100 g), iron and zinc (4.3–7.4 mg/100 g). Baru almond's protein did not show deficiency in essential amino acids and lysine was the first limiting amino acid in the proteins of the pequi almond and cerrado cashew nut. The baru almond showed a RNPR of 86%, similar to that of the cerrado cashew nut (78%), but higher than that of the peanut (72%) and of the pequi almond (54%). The PDCAAS value of the baru almond (91%) was the highest and cerrado cashew nut and peanut presented similar values of this index (82%), which were higher than that of the pequi almond (55%). The baru almond has the highest protein quality, but the cerrado cashew nut and peanut are sources of good quality protein, too. We recommend the inclusion of these exotic foods in healthy diets and in food industry, and the baru almond and cerrado cashew nut as sources of complementary protein.

Keywords: Native fruit; Brazilian Savanna; nut; almond; chemical composition; protein quality.

SUMÁRIO

	CAPÍTULO 1.....	9
1	INTRODUÇÃO.....	9
2	OBJETIVOS.....	12
2.1	OBJETIVO GERAL.....	12
2.3	OBJETIVOS ESPECÍFICOS.....	12
3	MATERIAL E MÉTODOS.....	13
3.1	OBTENÇÃO DAS SEMENTES COMESTÍVEIS E DA NOZ.....	13
3.2	PREPARO DAS AMOSTRAS.....	13
3.3	ANÁLISES QUÍMICAS.....	14
3.3.1	Composição centesimal.....	14
3.3.2	Perfil de aminoácidos.....	14
3.3.3	Análise de minerais.....	15
3.4	AVALIAÇÃO DA QUALIDADE PROTÉICA.....	16
3.4.1	Avaliação biológica.....	16
3.4.2	Determinação da qualidade protéica.....	17
3.5	ANÁLISES ESTATÍSTICAS.....	19
	REFERÊNCIAS.....	20
	CAPÍTULO 2 – Artigo científico.....	23
	CAPÍTULO 3 – Considerações finais.....	45
	ANEXOS.....	46

CAPÍTULO 1

1 INTRODUÇÃO

As nozes verdadeiras são frutas secas, espessas e muitas vezes contêm espinhos que recobrem sua semente. As mais conhecidas no mundo são: amêndoa, castanha-do-pará, castanha-de-caju, pistache, avelã, macadâmia, noz e castanha. Além das nozes verdadeiras, existem muitas sementes comestíveis com características semelhantes a elas, mas com classificação botânica diferente, como é o caso do amendoim (JUDD et al., 2002; VENKATACHALAN; SATHE, 2006).

Na flora nativa do Cerrado situada na região Central do Brasil, existem algumas espécies nativas como o barueiro, o pequizeiro e o cajueiro-do-cerrado que se destacam pela produção de seus frutos exóticos, de sabores marcantes e peculiares, como o baru (*Dipteryx alata* Vog.), o pequi (*Caryocar brasiliense* Camb.) e o caju-do-cerrado (*Anacardium othonianum* Rizz.), respectivamente. O baru possui semente, popularmente denominada de amêndoa ou castanha, que é comercializada em algumas cidades do Centro-Oeste. Por outro lado, a semente do pequi e o fruto do caju-do-cerrado são pouco explorados e, portanto, são descartados e pouco aproveitados na alimentação pela população do Cerrado.

O barueiro, destaca-se por seu porte, de 15 a 25 metros de altura e produz fruto, baru, de agosto a outubro, sendo estes de cor parda, contendo uma única semente oleaginosa comestível ou amêndoa (JUDD et al., 2002). O baru apresenta polpa com alta concentração de fibras insolúveis (cerca de 30%) e amêndoa com teores elevados de lipídios (cerca de 40%) e de proteínas (aproximadamente 30%), sendo esta de boa digestibilidade e que contém perfil de aminoácidos adequado às necessidades humanas (FERNANDES et al., 2010). Além dos macronutrientes, a amêndoa do baru possui teor considerável de minerais, com destaque para o cálcio, ferro, magnésio, potássio e zinco (FREITAS; NAVES, 2010; TAKEMOTO et al., 2001).

O pequizeiro é uma planta arbórea, de alto porte, de elevada frequência e ocorrência nas regiões do Cerrado. Seus frutos são constituídos de exocarpo ou

pericarpo, de coloração esverdeada ou marrom-esverdeada, mesocarpo externo, polpa branca com coloração parda acinzentada e mesocarpo interno, que constitui a porção comestível do fruto, possuindo coloração amarelada e separa-se facilmente do mesocarpo externo quando maduro. Além disso, é constituído pelo endocarpo, que é espinhoso, protege a semente ou amêndoa, que é também uma porção comestível do fruto (MELO JÚNIOR et al., 2004). Esta amêndoa possui elevado valor energético (aproximadamente 600 kcal/ 100 g), ressaltando seus teores de lipídios (cerca de 50%) e proteínas (acima de 20%) (LIMA et al., 2007).

Dentre as espécies de caju existentes no país, destaca-se, no estado de Goiás, o cajuzinho-do-cerrado, árvore mediana de 3 a 6m de altura (NAVES, 1999). O pseudofruto maduro, pedúnculo carnoso de coloração variando do amarelado ao vermelho vivo, é largamente consumido em razão das inúmeras possibilidades de utilização como sucos, doces e licores. O fruto verdadeiro, a castanha, pesa de 4 g a 13 g e é rica em lipídios. No entanto, observa-se que é pouco aproveitada para o consumo e são escassas as informações quanto ao seu valor nutritivo (CORREIA et al., 2008). No entanto, espera-se que esta castanha seja rica em proteína como a castanha-de-caju tradicional, a qual contém aproximadamente 20% deste nutriente (FREITAS; NAVES, 2010).

O amendoim (*Arachis hypogaea* L.) é uma semente oleaginosa e consiste na terceira fonte de proteína vegetal mais consumida em todo o mundo. Porém, este alimento apresenta deficiência em lisina (FERNANDES et al., 2010; SINGH; SINGH, 1991; VENKATACHALAM; SATHE, 2006). Assim como as amêndoas de baru e de pequi e a noz (castanha-de-caju-do-cerrado), o amendoim possui teores consideráveis de lipídios e proteínas (FERNANDES et al., 2010; LIMA et al., 2007; VENKATACHALAM; SATHE, 2006).

As nozes verdadeiras e as sementes comestíveis são, atualmente, bastante consumidas por serem fontes de substâncias com propriedades de alegação de saúde, ou compostos biologicamente ativos (FREITAS; NAVES, 2010; LÓPEZ-URIARTE et al., 2009). Além destes compostos biologicamente ativos, estes alimentos possuem quantidades elevadas de proteína. No entanto, existem poucos estudos que avaliaram o valor protéico de amêndoas e nozes, principalmente as nativas.

O valor protéico de alimentos é geralmente determinado pelo *Net Protein Ratio* (NPR) e pelo *Protein Digestibility-Corrected Amino Acid Score* (PDCAAS). O NPR é um método biológico, que avalia a eficiência de utilização da proteína usando ratos em crescimento (PELLETT; YOUNG, 1980). Este método, geralmente, subestima a qualidade protéica de alimentos de origem vegetal, uma vez que ratos em crescimento requerem maiores quantidades de alguns aminoácidos essenciais do que os humanos. O PDCAAS é um método recomendado pela FAO (1991) e IOM (2005) para avaliar a qualidade protéica, considerando tanto o perfil de aminoácidos essenciais (em comparação ao padrão de exigência humana) e a digestibilidade da proteína. Já foi elucidado que os valores de PDCAAS dos alimentos são geralmente maiores que os avaliados pelos índices biológicos. Assim, é importante avaliar a qualidade protéica por ambos os métodos - NPR, para estabelecer a relação entre a qualidade da proteína de diferentes alimentos, e PDCAAS, para estimar o valor protéico dos alimentos para o consumo humano (FRIEDMAN, 1996; SHAAFSMA, 2000).

O estudo do valor nutritivo e da qualidade protéica de frutos nativos do bioma Cerrado contribui para a utilização sustentável, a conservação e seleção das espécies promissoras, bem como a importância econômica destes alimentos. Assim, o objetivo deste estudo foi determinar a qualidade nutricional e o valor protéico de duas sementes comestíveis nativas (as amêndoas de baru e de pequi) e da castanha-de-caju-do-cerrado, e comparar estes valores com os do amendoim, que é amplamente consumido em todo mundo.

2 OBJETIVOS

2.1 OBJETIVO GERAL

Avaliar a qualidade nutricional e o valor protéico das amêndoas de baru (*Dipteryx alata* Vog.) e de pequi (*Caryocar brasiliense* Camb.) e da castanha-de-caju-do-cerrado (*Anacardium othonianum* Rizz.), de frutos oriundos de diferentes regiões do Cerrado, em relação ao amendoim (*Arachis hypogaea* L).

2.2 OBJETIVOS ESPECÍFICOS

- Analisar a composição centesimal, incluindo o teor de fibras alimentares totais (solúveis e insolúveis), das amêndoas de baru e de pequi, da castanha-de-caju-do-cerrado e do amendoim.
- Determinar o perfil de aminoácidos das amêndoas de baru e de pequi, da castanha-de-caju-do-cerrado e do amendoim.
- Quantificar o teor de minerais (Ca, Fe, K, Mg, Na, P, Se e Zn) das amêndoas de baru e de pequi, da castanha-de-caju-do-cerrado e do amendoim.
- Estimar o valor protéico das amêndoas de baru e de pequi, da castanha-de-caju-do-cerrado e do amendoim.
- Comparar a qualidade nutricional e o valor protéico das amêndoas de baru e de pequi e da castanha-de-caju-do-cerrado com a qualidade nutricional e o valor protéico do amendoim.

3 MATERIAL E MÉTODOS

3.1 OBTENÇÃO DAS SEMENTES COMESTÍVEIS E DA NOZ

Os frutos de baru foram coletados no mês de setembro de 2009, nas regiões Central (16° 24' 21"S e 49° 13' 08"O), Leste (15° 51' 07"S e 48° 57' 32"O) e Sul (16° 45' 43"S e 49° 41' 38"O) do estado de Goiás. Os pequis foram adquiridos na Ceasa-GO, provenientes de três regiões produtoras de pequi para comercialização (Minas Gerais, Tocantins e Goiás), durante o período de outubro a dezembro de 2009. O caju-do-cerrado foi coletado de plantas nativas da região Noroeste (15° 26' 45"S e 50° 21' 39"O) do estado de Goiás, durante o mês de setembro de 2009. Foi feita amostragem com número de frutos representativos para a pesquisa. O amendoim foi adquirido no mercado local de Goiânia.

Após o período de maturação, os frutos de pequi foram despolidos e secos em estufa a 60 °C por 30 horas (LIMA et al., 2007). As amêndoas de baru e de pequi foram extraídas com equipamento apropriado (guilhotina) (RABÊLO et al., 2008). A castanha-de-caju-do-cerrado foi extraída por método artesanal, conforme a seguir: o fruto do caju-do-cerrado foi disposto em um recipiente de alumínio perfurado e exposto ao fogo, com agitação constante para obter o escurecimento uniforme destes frutos e remover o líquido cáustico da castanha-de-caju. Posteriormente, a castanha-de-caju-do-cerrado foi decorticada com auxílio de faca de aço inoxidável.

3.2 PREPARO DAS AMOSTRAS

As amêndoas de baru e de pequi, a castanha-de-caju-do-cerrado e o amendoim foram torrados para inativar possíveis fatores antinutricionais (SGARBIERI, 1996) e para investigar a qualidade nutricional das amêndoas e da noz em sua forma usual de consumo. As amêndoas de baru e de pequi, e o amendoim foram torrados a 140 °C por 30 minutos (FERNANDES et al., 2010). A amêndoa do pequi e a castanha-de-caju-do-cerrado foram torrados a 130 °C por 30 minutos (RABÊLO et al., 2008), todos em forno elétrico. Posteriormente, as amêndoas e a noz foram moídas em multiprocessador doméstico e peneiradas (60

mesh) para análise química e para a elaboração das dietas experimentais oferecidas aos ratos durante o ensaio biológico.

3.3 ANÁLISES QUÍMICAS

3.3.1 Composição centesimal

A composição centesimal das amêndoas e da noz foi determinada, em três replicatas, por meio das análises de umidade e sólidos totais, conforme técnica descrita pelo Instituto Adolfo Lutz (2005); nitrogênio total, segundo o método de Kjeldahl e conversão em proteína bruta utilizando-se o fator 6,25 (AOAC, 1990); lipídios totais, extraídos por meio da técnica de Bligh e Dyer (1959) e posteriormente determinados por gravimetria; fibra alimentar solúvel e insolúvel, conforme técnica enzimica-gravimétrica descrita por Prosky et al. (1988) e as cinzas, por incineração em mufla a 550 °C (AOAC, 1990). Os carboidratos foram estimados por diferença, subtraindo-se de cem os valores obtidos para umidade, proteínas, lipídios, resíduo mineral fixo e fibra alimentar total. Todas as análises foram realizadas no Laboratório de Nutrição e Análise de Alimentos da Faculdade de Nutrição da UFG, exceto as análises de fibra alimentar, que foram realizadas no Laboratório de Pesquisa e Consultoria (LABM, Belo Horizonte). A partir dos dados da composição centesimal, foi estimado o valor energético (calórico) das amostras considerando-se os fatores de conversão de Atwater de 4, 4 e 9 para proteína, carboidrato e lipídio, respectivamente (MERRIL; WATT, 1973).

3.3.2 Perfil de aminoácidos

A composição de aminoácidos foi determinada no Centro de Química de Proteínas da Faculdade de Medicina de Ribeirão Preto da USP. As amostras foram submetidas à hidrólise ácida de proteínas e peptídeos, com solução aquosa de ácido clorídrico 6 N, bidestilado a 104 °C, contendo 0,1% de fenol (m/v) para a quantificação dos aminoácidos: lisina, histidina, arginina, ácido aspártico, treonina, serina, ácido glutâmico, prolina, glicina, alanina, cisteína, valina, metionina, isoleucina, leucina, tirosina e fenilalanina. Após a hidrólise ácida em solução, as

amostras foram secas em concentrador rotativo e ressuspensas em solução tampão de citrato de sódio 0,17 M, pH 2,2, contendo polietilenoglicol 400 a 15% (v/v) e tioglicol 0,4% (v/v) (MOORE; SPACKMAN; STEIN, 1958). Para a quantificação do aminoácido triptofano, as amostras foram submetidas à hidrólise alcalina com hidróxido de lítio 4 N, segundo técnica descrita por Lucas e Sotelo (1980). Após a hidrólise alcalina, as amostras foram neutralizadas com ácido ortofosfórico e filtradas em membrana de 0,45 micrômetros, para eliminar os precipitados. Foram adicionados à amostra, polietilenoglicol e solução tampão de citrato de sódio contendo tiodiglicol, para completar o volume em balão volumétrico. Em seguida, as amostras submetidas à hidrólise foram aplicadas em analisador automático de aminoácidos (Nicolas V, construído pelo Centro de Químicas de Proteínas, USP – Ribeirão Preto) e após eluição nas colunas e reação com ninidrina, os aminoácidos foram detectados colorimetricamente e quantificados. A partir dos resultados destas análises, foi estimado o escore de aminoácidos essenciais (EAE), que corresponde à proporção do aminoácido mais limitante (primeiro limitante) do alimento-teste em relação às necessidades de aminoácidos essenciais de crianças de dois a cinco anos de idade (pré-escolares) usadas como padrão de necessidade, de acordo com WHO/FAO/UNU (WHO, 2007).

3.3.3 Análise de minerais

As cinzas das amêndoas de baru e de pequi, da castanha-de-caju-do-cerrado e do amendoim obtidas por incineração a 550 °C foram solubilizadas com HCl concentrado (P.A). Os minerais (Ca, Fe, Mg, P e Zn) foram caracterizados e quantificados nas amostras por meio de espectrofotometria de absorção atômica (espectrofotômetro Varian®, modelo SpectrAA-200), utilizando-se os parâmetros instrumentais (lâmpada, comprimento de onda, corrente da lâmpada e largura da fenda) específicos para cada nutriente (AOAC, 1990). Os minerais Na e K foram analisados por fotometria de chama (fotômetro Analyser, modelo 910) (AOAC, 1990). As análises foram realizadas no Laboratório Nacional Agropecuário de Goiás (LANAGRO-GO) do Ministério da Agricultura, Pecuária e Abastecimento. O selênio também foi analisado por espectrofotometria de absorção atômica (espectrofotômetro HITACHI®, modelo Z-5000), por meio da geração de hidretos acoplados a cela de

quartzo, segundo procedimentos padronizados no Laboratório de Nutrição e Minerais da Faculdade de Ciências Farmacêuticas da USP (GONZAGA, 2002).

3.4 AVALIAÇÃO DA QUALIDADE PROTÉICA

3.4.1 Avaliação biológica

Para a realização do ensaio biológico foram utilizados 42 ratos machos, albinos, da linhagem Wistar, recém-desmamados, com pesos entre 50 e 70 g, provenientes do Laboratório Bioagri (Planaltina, Distrito Federal). Os ratos foram distribuídos aleatoriamente em grupos, segundo delineamento por blocos casualizados, contendo sete tratamentos com seis repetições. Os animais foram mantidos em gaiolas individuais de aço galvanizado durante dezessete dias, compreendendo um período preliminar de três dias para aclimação e 14 dias de experimento, sob condições ambientais padronizadas (ciclo de luz 12 h claro e 12 h escuro, temperatura média de 20 °C e umidade média de 60%, com trocas de ar freqüentes). A água destilada foi oferecida *ad libitum*. O consumo de dieta e o peso dos animais foram monitorados e registrados em planilhas em dias alternados. O ensaio biológico foi conduzido no Laboratório de Nutrição Experimental (Lanute) da Faculdade de Nutrição da UFG. Após o termino do experimento, os animais foram submetidos à eutanásia com éter etílico em recipiente fechado. Todos os procedimentos com os animais foram realizados de acordo com os princípios éticos preconizados pelo Colégio Brasileiro de Experimentação Animal (COBEA) (DE LUCA et al., 1990), e o protocolo experimental foi aprovado pelo Comitê de Ética da Universidade Federal de Goiás (Protocolo nº 153/08) (Anexo A).

Foram elaboradas sete dietas, segundo formulação básica preconizada por AIN-93G (REEVES; NIELSEN; FAHEY, 1993) contendo 10% de proteína: CAS7 (caseína com 7% de lipídios); CAS15 (caseína com 15% de lipídios); amêndoas de baru (AMB); amêndoas de pequi (AMP); castanha-de-caju-do-cerrado (CJC) e amendoim (AMD). Além dessas dietas, foi formulada uma dieta aprotéica (APO). A composição das dietas e os respectivos teores de proteínas e lipídios estão apresentados na Tabela 1. A amêndoa de baru e de pequi, a castanha-de-caju-do-cerrado e o amendoim não foram desengorduradas na confecção das dietas para conservar suas características naturais. Por isso, foi incluído o grupo CAS15

(caseína com 15% de lipídios) como controle interno do experimento, uma vez que as dietas com amêndoa de baru e de pequi, e amendoim ficaram hiperlipídicas, com teores de lipídios próximos a 15%, exceto a dieta com castanha-de-caju-do-cerrado.

Tabela 1. Ingredientes e composição química das dietas utilizadas no ensaio biológico

Componente (g/ 100g)	Dieta ¹						
	CAS 7	CAS 15	AMB	AMP	CJC	AMD	APO
Caseína	13,27	13,27	-	-	-	-	-
Amêndoa de baru	-	-	33,42	-	-	-	-
Amêndoa de pequi	-	-	-	33,73	-	-	-
Castanha-de-caju-do-cerrado	-	-	-	-	44,12	-	-
Amendoim	-	-	-	-	-	33,79	-
L-cistina	0,20	0,20	-	-	-	-	-
Óleo de soja	6,58	14,58	-	-	-	-	7,00
Celulose ²	5,00	5,00	0,52	4,26	3,37	2,30	5,00
Mistura salina	3,50	3,50	3,50	3,50	3,50	3,50	3,50
Bitartarato de colina	0,25	0,25	0,25	0,25	0,25	0,25	0,25
Mistura vitamínica	1,00	1,00	1,00	1,00	1,00	1,00	1,00
Amido de milho	70,20	62,20	61,31	57,26	47,76	59,16	83,25
% Proteína ³	10,39	10,67	9,24	9,94	9,93	10,95	0,45
% Lipídios ³	6,7	15,89	14,36	17,24	21,08	15,63	15,82
VET ⁴	412,50	452,50	452,00	459,10	485,40	458,20	-

¹ Formulação de acordo com AIN-93G (REEVES; NIELSEN; FAHEY, 1993), com redução dos teores de proteína para 10%. CAS7: caseína com 7% de lipídios (referência); CAS15: caseína com 15% de lipídios (controle); AMB: amêndoa de baru torrada; AMP: amêndoa de pequi torrada; CJC: castanha-de-caju-do-cerrado torrada; AMD: amendoim torrado; APO: aprotéica.

² Celulose: adicionada de acordo com o teor de fibra desses alimentos - amêndoa de baru (TAKEMOTO et al., 2001); amendoim e castanha-de-caju (NEPA, 2006).

³ Resultados das análises químicas das dietas formuladas.

⁴ Valor Energético Total das dietas, calculado por meio dos fatores de conversão de 4, 4 e 9 para proteínas, carboidratos e lipídios, respectivamente (MERRIL; WATT, 1973).

3.4.2 Determinação da qualidade protéica

A eficiência protéica e a qualidade protéica das amêndoas de baru e de pequi e da castanha-de-caju-do-cerrado e do amendoim foram avaliadas pelos seguintes métodos: NPR (Net Protein Ratio), RNPR (Relative Net Protein Ratio) e PDCAAS (Protein Digestibility-Corrected Amino Acid Score).

Para avaliar a eficiência protéica das amêndoas de baru e de pequi, da castanha-de-caju-do-cerrado e do amendoim, em manter e aumentar o peso de animais experimentais, utilizou-se o método NPR (Net Protein Ratio) no 14º dia do experimento, segundo Pellett e Young (1980). Levou-se em consideração o ganho de peso do grupo teste mais a perda de peso do grupo com dieta aprotéica, em relação ao consumo de proteína do grupo teste, correspondendo à fórmula:

$$\text{NPR} = \frac{\text{Ganho de peso (grupo teste)} + \text{Perda de peso (grupo aprotéico)}}{\text{Proteína ingerida (grupo teste)}}$$

A partir dos dados de NPR, foram calculados os valores de RNPR, que mede a porcentagem do NPR do grupo teste em relação à proteína de referência (caseína).

A qualidade protéica foi também avaliada pelo método PDCAAS (Protein Digestibility-Corrected Amino Acid Score), preconizado pela FAO (1991). O PDCAAS considera dois parâmetros na avaliação da qualidade protéica: a digestibilidade e a capacidade da proteína de suprir as necessidades de aminoácidos essenciais de humanos. A digestibilidade é avaliada pelo quociente do nitrogênio absorvido pelo nitrogênio ingerido na dieta, expresso em porcentagem. Para a determinação do nitrogênio fecal, as dietas foram marcadas no 7º dia de experimento com índigo carmim P.A. (50 mg/ 100 g de dieta), no 8º dia de experimento coletou-se apenas as fezes marcadas com carmim, e do 9º ao 14º dia de experimento todas as fezes foram coletadas, acondicionadas em recipientes individuais para cada animal, e mantidas sob condições de refrigeração. Durante todos os dias de coleta de fezes, estas foram pesadas e o peso registrado em planilha. Posteriormente ao período de coleta, as fezes foram secas em estufa a 100°C, por 24 horas, depois resfriadas em dessecador, pesadas e então trituradas para determinação do teor de nitrogênio, pelo método de micro-Kjeldahl (AOAC, 1990). A digestibilidade verdadeira (Dv) das fontes protéicas foi determinada pela obtenção da quantidade de nitrogênio ingerido pelos ratos (I), a quantidade de nitrogênio excretado nas fezes pelo grupo de ratos com dieta protéica (F), e a quantidade de nitrogênio fecal metabólico (endógeno) excretado nas fezes pelo grupo de ratos com dieta aprotéica (Fe). Assim, a digestibilidade foi calculada pela seguinte fórmula, conforme FAO (1991):

$$\text{DV} = \frac{\text{I} - (\text{F} - \text{Fe})}{\text{I}} \times 100$$

A partir desses dados, o PDCAAS foi determinado por meio do produto do EAE (estimado conforme item 3.4.2) e Dv da proteína, isto é:

$$\text{PDCAAS (\%)} = \frac{\text{EAE} \times \text{DV}}{100}$$

3.5 ANÁLISES ESTATÍSTICAS

Os resultados da composição centesimal e do ensaio biológico foram submetidos à análise de variância e teste para comparação de médias (Tukey a 5% de probabilidade). Foi feita análise de regressão linear do peso dos animais em relação ao tempo de experimento. Os cálculos estatísticos foram efetuados com auxílio do programa Excel-versão 2007 e do programa Statistica - Stat Soft Inc., versão 7.0, 2004, Tulsa, EUA.

REFERÊNCIAS

AOAC - ASSOCIATION OF OFFICIAL ANALYTICAL CHEMISTS. **Official methods of analysis**. 15. ed. Arlington: AOAC, 1990.

BLIGH, E. G.; DYER, W. J. A rapid method of total lipid extraction and purification. **Canadian Journal of Biochemistry and Physiology**, Toronto, v.37, n.8, p.911-917, 1959.

CORREIA, G. C.; NAVES, R. V.; ROCHA, M. R.; CHAVES, L. J.; BORGES, J. D. Determinações físicas em frutos e sementes de baru (*Dipteryx alata* Vog., cajuzinho (*Anacardium othonianum* Rizz.) e pequi (*Caryocar brasiliense* Camb. **Bioscience Journal**, Uberlândia, v.24, n.4, p. 42-47, 2008.

DE LUCA, R. R.; ALEXANDRE, S. R.; MARQUES, T.; SOUZA, N. L.; MERUSSE, J. L. B.; NEVES, S. P. (Ed.). **Manual para técnicos em bioterismo**. 2. ed. São Paulo: Winner Graph, 1996.

FAO - FOOD AND AGRICULTURE ORGANIZATION OF THE UNITED NATIONS. Report of a Joint FAO/WHO Expert Consultation. **Protein quality evaluation**. Rome: FAO, 1991. 66p. (FAO Food and Nutrition Paper, 51).

FERNANDES, D. C.; FREITAS, J. B.; CZEDER, L. P.; NAVES, M. M. V. Nutritional composition and protein value of the baru (*Dipteryx alata* Vog.) almond from the Brazilian Savanna. **Journal of the Science of Food and Agriculture**, Oxford, v.90, n. 9, p. 1650-1655, 2010.

FREITAS, J. B.; NAVES, M. M. V. Composição química de nozes e sementes comestíveis e sua relação com a nutrição e saúde. **Revista de Nutrição**, Campinas, v.23, n.2, p.269-279, 2010.

FRIEDMAN, M. Nutritional value of proteins from different food sources. A review. **Journal of Agriculture and Food Chemistry**, Washington, v. 44, n. 1, p.6–29, 1996.

GONZAGA, I. B. **Avaliação nutricional relativa ao selênio em crianças com dieta enriquecida de castanha-do-brasil (*Bertholletia excelsa*, L.)**. 2002. 161 f. Tese (Doutorado em Ciência dos Alimentos) – Faculdade de Ciências Farmacêuticas, Universidade de São Paulo, São Paulo, 2002.

INSTITUTO ADOLFO LUTZ. **Normas analíticas do Instituto Adolfo Lutz: métodos físico-químicos para análise de alimentos**. 4. ed. Brasília, DF: Ministério da Saúde, Agência Nacional de Vigilância Sanitária, 2005. 1018p.

IOM - INSTITUTE OF MEDICINE. FOOD AND NUTRITION BOARD (FNB). Protein and amino acids. In: IOM. FNB. **Dietary references intakes for energy, carbohydrate, fiber, fat, fatty acids, cholesterol, protein, and amino acids**. Washington DC: USA National Academies, 2005. cap. 10, p. 589-768.

JUDD, W. S.; CAMPBELL, C. S.; KELLOGG, E. A.; STEVENS, P. F.; DONOGHUE, M. J. Taxonomic evidence: structural and biochemical characters. In: _____. **Plant systematics: a phylogenetic approach**. 2. ed. Massachusetts: Sunderland, 2002. cap. 3. p. 55-104.

LIMA, A.; SILVA, A. M. O.; TRINDADE, R. A.; TORRES, R. P.; MANCINI-FLHO, J. Composição química e compostos bioativos presentes na polpa e na amêndoa de pequi (*Caryocar brasiliense* Camb.). **Revista Brasileira de Fruticultura**, Jaboticabal, v.29, n.3, p.695-698, 2007.

LÓPEZ-URIARTE, P.; BULLÓ, M.; CASAS-AGUSTENCH, P.; BABIO, N.; SALAS-SALVADO, J. Nuts and oxidation: a systematic review. **Nutrition Reviews**, New York, v. 67, n. 9, p. 497–508, 2009.

LUCAS, B.; SOTELO, A. Effect of different alkalies, temperatures and hydrolises times on tryptophan determination of pure proteins and foods. **Analytical Biochemistry**, Washington, v.109, n.1, p. 192-197, 1980.

MELO JÚNIOR, A. F.; CARVALHO, D.; PÓVOA, J. S. R.; BEARZOTI, E. Estrutura genética de populações naturais de pequizeiro (*Caryocar brasiliense* Camb.). **Scientia Forestalis**, Piracicaba, n.66, p. 56-65, 2004.

MERRIL, A. L.; WATT, B. K. **Energy value of foods: basis and derivation**. Washington: United States Department of Agriculture, 1973. 105p.

MOORE, S.; SPACKMAN, D. H.; STEIN, W. H. Chromatography of amino acids on sulfonated polystyrene resins. **Analytical Chemistry**, Washington, v.30, n.7, p. 1185-1190, 1958.

NAVES, R. V. **Espécies frutíferas dos cerrados de Goiás: caracterização e influências do clima e dos solos**. Goiânia, 1999. 202 p. Tese (Doutorado em Agronomia) - Escola de Agronomia e Engenharia de Alimentos, Universidade Federal de Goiás, Goiânia, 1999.

NEPA - NÚCLEO DE ESTUDOS E PESQUISAS EM ALIMENTAÇÃO. **Tabela brasileira de composição de alimentos**. 2. ed. Campinas: Nepa-Unicamp, 2006.113p.

PELLETT, P. L.; YOUNG, V. R. **Nutritional evaluation of protein foods**. Tokyo: The United Nations University, 1980. 154p.

PROSKY, L.; ASP, N.; SCHWEIZER, T. F.; DEVRIES, J. W.; FURDA, I. Determination of insoluble, soluble, and total dietary fiber in foods and food products: interlaboratory study. **Journal of the Association of Official Analytical Chemists**, Arlington, v.71, n.5, p. 1017-1023, 1988.

RABÊLO, A. M. S.; TORRES, M. C. L.; GERALDINE, R. M.; SILVEIRA, M. F. A. Extração, secagem e torrefação da amêndoa do pequi (*Caryocar brasiliense* Camb.). **Ciência e Tecnologia de Alimentos**, Campinas, v.28, n.4, p. 868-871, 2008.

REEVES, P. G.; NIELSON, F. H.; FAHEY JR., G., C. AIN-93 purified diets for laboratory rodents: final report of the American institute of Nutrition ad hoc writing committee on the reformulation of the AIN-76A rodent diet. **Journal of Nutrition**, Philadelphia, v.123, n. 11I, p. 1939-1951, 1993.

SCHAAFSMA, G. The protein digestibility-corrected amino acid score. **Journal of Nutrition**, Philadelphia, v. 130, n. 7, p.1865S–1867S, 2000.

SGARBIERI, V. C. Propriedades nutricionais das proteínas. In:_____. **Proteínas em alimentos protéicos: propriedades, degradações, modificações**. São Paulo: Varela, 1996. cap. 4, p. 337-386.

SINGH, B.; SINGH, U. Peanut as a source of protein for human foods. **Plant Foods for Human Nutrition**, Dordrecht, v. 41, n. 2, p. 165–177, 1991.

TAKEMOTO, E.; OKADA, I. A.; GARBELOTTI, M. L.; TAVARES, M.; AUEDPIMENTEL, S. Composição química da semente e do óleo de baru (*Dipteryx alata* Vog.) nativo do município de Pirenópolis, estado de Goiás. **Revista do Instituto Adolfo Lutz**, São Paulo, v. 60, n. 2, p. 113-117, 2001.

VENKATACHALAM, M.; SATHE, S. K. Chemical composition of selected edible nut seeds. **Journal of Agricultural and Food Chemistry**, Washington, v.54, n.13, p. 4705-4714, 2006.

WHO - WORLD HEALTH ORGANIZATION. Report of a Joint WHO/FAO/UNU Expert Consultation. **Protein and amino acid requirements in human nutrition**. Geneva: WHO, 2007. 265 p. (WHO Technical Report Series, 935).

CAPÍTULO 2 - ARTIGO CIENTÍFICO

O artigo científico foi submetido e aprovado para publicação no periódico Food Research International, conforme a seguinte referência:

SOUSA, A. G. O.; FERNANDES, D. C.; ALVES, A. M.; FREITAS, J. B.; NAVES, M. M. V. Nutritional quality and protein value of exotic almonds and nut from the Brazilian Savanna compared to peanut. **Food Research International**, Essex, v. 44, n. 4, 2011. Disponível em <http://dx.doi.org/10.1016/j.foodres.2011.02.013>.

Este periódico possui qualificação Qualis B1 (Anexo B) na área de Medicina II da Capes e as normas para elaboração do artigo estão descritas no Anexo C.

Nutritional quality and protein value of exotic almonds and nut from the Brazilian Savanna compared to peanut

Amanda Goulart de Oliveira Sousa^a, Daniela Canuto Fernandes^b, Aline Medeiros Alves^a,
Jullyana Borges de Freitas^c, Maria Margareth Veloso Naves^{d*}

^aFaculty of Nutrition, Federal University of Goiás (Universidade Federal de Goiás – UFG), Goiânia, GO, Brazil

^bFaculty of Agronomy and Food Engineering, Federal University of Goiás, Goiânia, GO, Brazil

^cFederal Institute of Education, Science and Technology, Goiânia, GO, Brazil

^dExperimental Nutrition Laboratory, Faculty of Nutrition, Federal University of Goiás, Goiânia, GO, Brazil

*Corresponding author: Maria Margareth V. Naves, Experimental Nutrition Laboratory, Faculty of Nutrition, UFG, Rua 227, Quadra 68, 74605-080, Goiânia, GO, Brazil. Phone: +55 (62) 3209-6270/214. Fax: +55 (62) 3209-6273. E-mail: mnaves@fanut.ufg.br

Abstract

The aim of this study was to determine the nutritional quality and protein value of the baru almond, pequi almond, and cerrado cashew nut, native fruits from the Brazilian Savanna, compared to the peanut. We determined the proximate chemical composition, mineral content, and amino acid profile. A biological assay was carried out to assess the protein value, by net protein ratio (NPR), relative net protein ratio (RNPR), and protein digestibility-corrected amino acid score (PDCAAS) indexes. We found that the exotic almonds and the nut are rich in proteins (22.7–29.9 g/100 g), lipids (41.9–50.0 g/100 g), fibres (baru and pequi almonds, around 10.0 g/100 g), iron and zinc (4.3–7.4 mg/100 g). Baru almond's protein did not show deficiency in essential amino acids and lysine was the first limiting amino acid in the proteins of the pequi almond and cerrado cashew nut. The baru almond showed a RNPR of 86%, similar to that of the cerrado cashew nut (78%), but higher than that of the peanut (72%) and of the pequi almond (54%). The PDCAAS value of the baru almond (91%) was highest and cerrado cashew nut and peanut presented similar values of this index (82%), which were higher than that of the pequi almond (55%). The baru almond has the highest protein quality, but the cerrado cashew nut and peanut are sources of good quality protein, too. We recommend the inclusion of these exotic foods in healthy diets and in food industry, and the baru almond and cerrado cashew nut as sources of complementary protein.

Keywords: Native fruit; Brazilian Savanna; Nut; Almond; Chemical composition; Protein quality

1. Introduction

A true nut is a dry fruit often covered by a shell with thorns that protects the seed. The most popular nuts across the world are the Brazil nut, cashew nut, pistachio, walnut, macadamia, nut, and chestnut. In addition to the true nuts, there exist other seeds with similar morphological and sensory qualities but with a different botanical classification, such as the peanut (Judd, Campbell, Kellogg, Stevens, & Donoghue, 2002; Venkatachalan & Sathe, 2006).

Several tree species flourish in the native flora of the Cerrado region of central Brazil (Savanna). These include the ‘baru’ tree (*Dipteryx alata* Vog.), the ‘pequi’ tree (*Caryocar brasiliense* Camb.), and the ‘caju-do-cerrado’ tree (*Anacardium othonianum* Rizz.). All these 3 species give exotic-tasting fruits: the baru, pequi, and cerrado cashew, respectively.

The baru tree (Fabaceae family) produces fruits from August to October. The brown fruits contain a single edible oleaginous seed, commonly named almond (Fig. 1). The baru almond contains high levels of lipids (around 40%) and proteins (approximately 30%) with good digestibility and amino acid profile (Fernandes, Freitas, Czedler, & Naves, 2010). Moreover, the baru almond has a high content of minerals, particularly calcium, iron, magnesium, potassium, and zinc (Freitas & Naves, 2010; Takemoto, Okada, Garbelotti, Tavares & Aued-Pimentel, 2001).

The pequi tree is common in the Cerrado regions (Savanna) and its fruit consists of a greenish or greenish-brown exocarpus or pericarpus, an external mesocarpus (white pulp), and an internal yellow–dark orange mesocarpus (the fruit’s edible part). Furthermore, a thorny endocarpus protects the edible seed, also named almond (Melo Júnior, Carvalho, Póvoa, & Bearzoti, 2004) (Fig. 1). The energy value of almond is high (approximately 600 kcal/100 g), with considerably high lipid (about 50%) and protein contents (over 20%) (Lima, Silva, Trindade, Torres, & Mancini-Filho, 2007).

The ‘cerrado cashew nut’ is an important Brazilian cashew specie native from the Savanna biome of the Goiás state. The mature pseudo-fruit (fleshy peduncle) has a colour that varies from yellowish to bright red and is widely used in juices, sweets, and liqueurs. In turn, the true fruit or nut (weight 4–13 g) is rarely consumed, and its nutritional value is not yet known (Correia, Naves, Rocha, Chaves, & Borges, 2008) (Fig. 1).

The peanut (*Arachis hypogaea* L.), also known as groundnut, earthnut, monkeynut or ground bean, is the world’s third most important source of vegetable protein, yet lysine-

deficient (Fernandes, Freitas, Czeder, & Naves, 2010; Singh & Singh, 1991; Venkatachalam & Sathe, 2006). The peanut has considerably high amounts of lipids and proteins, which are similar to those of the baru and pequi almonds (Fernandes et al., 2010; Lima et al., 2007; Venkatachalam & Sathe, 2006).

True nuts and edible seeds are widely consumed as sources of health-promoting substances and of functional or biological active compounds (Freitas & Naves, 2010; López-Uriarte, Bulló, Casas-Agustench, Babio, & Salas-Salvado, 2009). In addition to biological active compounds, these foods have high amounts of protein. However, few studies have determined the protein value of nuts and almonds, especially of the exotic ones. Therefore, research is necessary to generate comprehensive knowledge about the protein value of native almonds and nuts.

The protein value of food is usually determined by net protein ratio (NPR) and protein digestibility-corrected amino acid score (PDCAAS) indexes. NPR is a biological method that assesses the efficiency of protein utilisation by using growing rats (Pellett & Young, 1980). This method often underestimates the protein quality of plant foods, since growing rats require higher amounts of certain essential amino acids than humans. PDCAAS is a method recommended by FAO (1991) and IOM (2005) to evaluate the protein quality considering both essential amino acid profile (in comparison with the human requirement pattern) and protein digestibility. It has been previously shown that the PDCAAS values of foods are generally higher than those evaluated by biological indexes. Thus, it is important to evaluate the protein quality by both methods – NPR, to establish the relative protein quality among different food sources, and PDCAAS, to estimate more reliably the protein value of food for human consumption (Friedman, 1996; Shaafsma, 2000).

The study of the protein and nutritive value of native fruits from the Savanna biome contributes to the sustainable use, conservation, and selection of the promising species as well as to the economic importance of these foods. Thus, the purpose of this study was to determine the nutritional quality and protein value of 2 native edible seeds (the baru and pequi almonds) and of cerrado cashew nut and to compare these values to those of the peanut, which is widely consumed worldwide.

2. Material and methods

2.1 Fruit collection

Baru and pequi fruits were collected from representative regions of their productivity in the Brazilian Savanna. The cerrado cashew fruit was collected in only one region (Northwest) of the Goiás state (Brazil) because this region had the highest productivity in the year of collection. Peanuts (variety HPS) were purchased from the local market of Goiânia, Goiás state, Brazil.

2.2 Sample preparation procedures

After maturation period, the pequi fruits were pulped and dried in an oven at 60°C for 30 h. The baru and pequi almonds (Fig. 1) were extracted using a guillotine. The cerrado cashew nut (Fig. 1) was extracted manually, as follows: the fruit of the cerrado cashew was directly exposed to fire in a perforated steel container to remove the corrosive liquid of the cashew nut; subsequently, the cerrado cashew nut was decorticated using a stainless steel knife. The baru and pequi almonds and the cerrado cashew nut were roasted to inactivate the possible anti-nutritional factors (Togashi & Sgarbieri, 1994) and to investigate the nutritional quality of the almonds and of the nut as usually consumed. The baru almond and the peanuts were roasted at 140°C for 30 min (Fernandes et al., 2010) and the pequi almond and cerrado cashew nut at 130°C for 30 min (Rabêlo, Torres, Geraldine, & Silveira, 2008) in an electric oven. Thereafter, the almonds and the nut were milled with a blender and sieved through a 60-mesh for chemical analysis and for use in the experimental diets.

2.3 Chemical characterization of the exotic almonds and the nut

2.3.1 Chemical composition

The following chemical composition parameters were determined in triplicate: moisture (AOAC, 1990); nitrogen content by the micro-Kjeldahl method (AOAC, 1990) by using a conversion factor of 6.25 (FAO, 1970); total lipids by extraction with chloroform and methanol (Bligh & Dyer, 1959); ash at 550°C (AOAC, 1990); and total dietary fibres (soluble and insoluble), according to the enzymatic-gravimetric technique (Prosky, Asp, Schweizer, Devries, & Furda, 1988). The carbohydrate content was estimated by subtracting from 100 the values obtained for moisture, protein, lipid, ash, and total dietary fibre contents. The energy value was estimated by using Atwater conversion factors of 4 kcal (proteins and carbohydrates), and 9 kcal (lipids) (Merril & Watt, 1973).

2.3.2 Minerals

Minerals (Ca, Fe, K, Mg, Na, P, and Zn) were characterised and quantified, in triplicate, by atomic absorption spectrophotometry (Perkin Elmer Analyst-200 spectrometer). Samples (30 g) were incinerated and then dissolved with concentrated hydrochloric acid (analytical grade). Specific instrumental parameters (lamp, wavelength, lamp current, and slit width) were used for each mineral (AOAC, 1990). Selenium was analysed by atomic absorption spectrophotometry (HITACHI®, model Z-5000, Tokyo) with generation of hydrides in a quartz tube.

2.3.3 Amino acid profile

The amino acid profile analysis of the exotic almonds and the nut were performed in duplicate. The quantification of amino acids (with the exception of tryptophan) was performed by acid hydrolysis of proteins and peptides (Moore, Spackman, & Stein, 1958). Tryptophan was quantified in samples treated by alkaline hydrolysis (Lucas & Sotelo, 1980). Then, samples were placed in an automatic amino acid analyser (Nicolas V, Protein Chemistry Centre, University of São Paulo, Ribeirão Preto, Brazil). After elution in the column and reaction with ninhydrin, the amino acids were quantified by a colorimetric assay.

The results of this analysis were used to estimate the amino acid score (AAS) by using the following formula (WHO, 2007): (mg of amino acid in 1 g of test protein/mg of amino acid in requirement pattern) $\times$ 100.

2.4 Evaluation of the protein value

2.4.1 Biological assay

The experiment was carried out with 42 weaned male Wistar rats (21–23 days old) from Bioagri Laboratories (Planaltina, Federal District, Brazil). The rats were randomly divided into 7 groups of 6 animals each, and kept in individual cages under standard environmental conditions (temperature $23 \pm 2^\circ\text{C}$, relative humidity 50–60%, 12/12 h light/dark cycle). All procedures with animals were conducted in accordance with the Guide for the Care and Use of Laboratory Animals (NRC, 1996) and were approved by the Research Ethics Committee of the Federal University of Goiás (Protocol no. 153/2008).

The animals were fed for 17 days (3 days of acclimatisation and 14 days of experiment) with experimental diets. The diets were formulated on the basis of AIN-93G

formula (Reeves, Nielsen, & Fahey Jr., 1993) with modification in the protein content (10%). The experimental groups were divided into 2 casein diets (casein diet with 7% lipids [reference group] and casein diet with 15% lipids [control group]), 4 experimental diets (baru almond, pequi almond, cerrado cashew nut, and peanut, with approximately 15% lipids), and a protein-free diet. The milled almond and nut used in the diets were not defatted in order to preserve their natural characteristics. The ingredients and chemical composition of these diets are shown in Table 1.

The body weight and food intake (food offered – food wasted) in all experimental groups were monitored on alternate days. Potable water was provided *ad libitum*. At the end of the experiment, the animals were weighed and euthanized with ethyl ether in a closed container.

2.4.2 Estimative of biological indexes

The protein values of the native fruits and of the peanut were estimated using the net protein ratio (NPR) and the protein digestibility-corrected amino acid score (PDCAAS) indexes. NPR and RNPR (relative NPR) were calculated as follows: $NPR = [\text{weight gain of test group (g)} + \text{weight loss of protein-free group (g)}] / \text{protein intake of test group (g)}$; $RNPR = [NPR \text{ test group} / NPR \text{ reference group} \times 100]$ (Pellett & Young, 1980). True protein digestibility (%) was determined as recommended by the Food and Agriculture Organization (FAO) (FAO, 1991) for *in vivo* test. Faeces were marked and collected during the second week of the experiment, and ground for nitrogen analysis. The amount of nitrogen consumed by the animals (I), the amount of nitrogen excreted in the faeces of animals fed with a protein diet (F), and the amount of nitrogen excreted in the faeces of animals fed with a protein-free diet (endogenous nitrogen – Fk) were determined. True digestibility was calculated using the formula: $[I - (F - Fk) / I] \times 100$. PDCAAS was estimated as follows: $[AAS \text{ of test protein (\%)} \times \text{true digestibility of test protein (\%)}] / 100$ (FAO, 1991).

2.5 Statistical analysis

The data are presented as mean $\pm$ standard deviation. Analysis of variance and Tukey's mean comparison test were used to compare the data about chemical composition and data from the biological assay. STATISTICA version 7.0 (StatSoft, Inc., Tulsa, OK,

USA, 2004) was used for statistical analyses. Differences were considered significant when the *P* value was <0.05.

3. Results and Discussion

The chemical composition and the energy values of the exotic almonds and nut (baru almond, pequi almond, and cerrado cashew nut) and peanuts are shown in Table 2. We found that the baru almond, pequi almond, and peanut have high protein contents. The exotic almonds and the nut contain high lipid content (40–50 g/100 g) and elevated energy value. In addition, these fruits contained high amounts of dietary fibres (>5 g/100 g), except the cerrado cashew nut (Table 2).

The protein and lipid contents and the energy value of the roasted baru almond are similar to those reported in the literature (26 g/100 g, 42 g/100 g and 535 kcal/100 g, respectively) (Fernandes et al., 2010). This edible seed presented a high content of dietary fibres (mainly insoluble dietary fibres), in accordance with the literature (13.4 g/100 g) (Takemoto et al., 2001). The roasted pequi almond showed the highest lipid content (50%), similar to that reported in the literature (51%) (Lima et al., 2007). It also presented the highest level of total dietary fibres, soluble fibres and ash. Cerrado cashew nut presented the highest energy density (about 600 kcal/100 g). There is no study in the literature about the chemical composition of the cerrado cashew nut. However, its chemical composition and energy value were similar to those of the traditional cashew nut (Venkatachalam & Sathe, 2006). The peanut showed similar chemical composition and energy value (Table 2) to those reported in the literature for this edible seed (Fernandes et al., 2010; Venkatachalam & Sathe, 2006).

The exotic almonds and the nut showed high concentrations of iron, potassium, magnesium and zinc (Table 3). On the other hand, these foods had low sodium content. Notably, the roasted baru almond showed the highest concentrations of calcium, and the other exotic fruits presented greater levels of calcium than that of the peanut. The roasted pequi almond showed the highest concentrations of magnesium, selenium, and zinc, the latter meeting 67% of the dietary reference intake for adults (IOM, 2006). The amount of zinc present in the roasted pequi almond was higher than that in any almond or nut reported in the literature (Freitas & Naves, 2010). Considering the nutritional importance of this mineral as an antioxidant, and its limited content in plant foods, the pequi almond can be considered as a good source of zinc. Peanut had the highest selenium content compared to the other exotic

fruits (Table 3). There is no published data about the selenium content of the exotic almonds and nut (Freitas & Naves, 2010). The amount of selenium in cerrado cashew nut (Table 3) is comparable to that in the traditional cashew nut ($3.0 \mu\text{g}/100 \text{ g}$) (Fagbemi, 2008).

A deeper knowledge of the mineral composition and bioavailability of the exotic almonds and nuts native to the Brazilian Savanna is useful for the prevention of mineral deficiencies, especially for communities with limited access to quality diet.

The amino acid composition of the baru almond, pequi almond, cerrado cashew nut and peanut are shown in Table 4. In general, cerrado cashew nut and exotic almonds have higher concentrations of sulphur amino acids than those of the common bean (FAO, 1991), a staple food of the Brazilian diet. According to the requirement pattern of essential amino acids (WHO, 2007), the proteins of the baru almond met 100% of the nutritional requirements. Therefore, the baru almond studied has more sulphur-containing amino acids (Table 4) than that of reported in the literature ($22 \text{ mg amino acid/g protein}$) (Fernandes et al., 2010).

The limiting amino acids of the pequi almond were lysine (first limiting amino acid), isoleucine, and threonine (Table 4). Lysine was the limiting amino acid of the cerrado cashew nut, in agreement to what's reported in the literature for the traditional cashew nut (Venkatachalan & Sathe, 2006). According to literature, lysine is the first limiting amino acid in some other nuts such as the Brazil nuts, traditional cashew nuts, hazelnuts, pistachios, almonds, and macadamia nuts (Ruggeri, Cappelloni, Gambelli, & Carnovale, 1998; Venkatachalan & Sathe, 2006). There are no reports on the amino acid profile of pequi almond and cerrado cashew nut. Peanut showed valine as the first, and lysine as the second limiting amino acid (Table 4), as previously reported (Venkatachalan & Sathe, 2006).

The total essential amino acid content of baru almond was higher than those of the other fruits, including the peanut. In turn, pequi almond showed the lowest level of essential amino acids, mainly because of its low lysine content (Table 4).

The body weight profiles of the experimental groups are shown in Fig. 2. The body-weight gain of rats fed with the baru almond and cerrado cashew diets was similar to that of the peanut diet-fed rats, but lower than that of the casein diet-fed rats (Fig. 2, Table 5). All groups of animals, except protein-free group, showed a positive linear trend of body weight evolution over time, yet the animals fed with pequi almond diet showed more moderate weight gain ($P < 0.05$; Fig. 2). The growth rates of the animals are represented by the 'b' value in the linear regression equation, and there are no significant differences among baru almond, cerrado cashew nut, and peanut groups ('b' values around 3, $P > 0.05$). Despite the

different energy values of these diets, the growth rate of the 2 groups of animals fed with casein diets was similar ('b' values around 5, $P > 0.05$), probably because the control group showed a compensatory reduction in its food intake in comparison with the reference group (Table 5).

The food and protein intakes of the animals are shown in Table 5. Food intakes of the rats fed with the baru almond, cerrado cashew nut, and peanut diets were similar. Food intake of the pequi almond group was lower than that of the other groups. It should be pointed that the essential amino acid composition of a diet influences the food intake, mainly because of an imbalance in the essential amino acid content (Harper, Benevenga, & Wohlhueter, 1970). Furthermore, the pequi almond has a strongly marked flavour, which can also be contributed to the low food intake of this group. The protein intake of the rats fed with the baru and peanut diets were similar to that of the cerrado cashew nut group, and the pequi almond group presented the lowest protein intake, because of its lowest food intake (Table 5).

The proteins of the exotic fruits had similar true digestibility (Table 5). The peanut's protein showed the highest true digestibility among the experimental groups. This value was in agreement with a previous report (92%) (Fernandes et al., 2010). On the other hand, the true digestibility of the baru almond's protein was higher than the values previously reported, of 80% (Fernandes et al., 2010) and 66% (Togashi & Sgarbieri, 1995). There are no reports on the protein digestibility of the pequi almond and of the cerrado cashew nut. According to the FAO (1970), the digestibility of the protein of the traditional cashew nut is 85%, comparable to the digestibility of the cerrado cashew nut's protein (Table 5).

According to the RNPR values, the baru almond presented protein value similar to that of the cerrado cashew nut and higher than that of the peanut (Table 5). Besides, the baru almond showed the highest PDCAAS among the experimental groups, which was higher than the value of 73%, reported in the literature (Fernandes et al., 2010). The PDCAAS of the peanut was very close to that observed for the cerrado cashew nut (Table 5). In turn, the RNPR and PDCAAS of the pequi almond were lower than the other groups, probably because of its poor amino acid profile (Table 4). These results indicate that these exotic fruits, except the pequi almond, contain good quality proteins (PDCAAS around 85%). The data of this study reinforce that the NPR index is more sensible to evince slightly differences in protein quality than PDCAAS index (Table 5), but the PDCAAS method is the most appropriate to estimate the food protein value for humans (Friedman, 1996; IOM, 2005; Schaafsma, 2000).

Our findings contribute to public awareness of the health benefits of exotic fruits consumption. Beside, the protein values of the pequi almond and cerrado cashew nut have not been determined so far, and these informations can be useful to the preservation of these exotic species (Fig. 1). The study of exotic edible seeds is necessary because these foods have nutritional values similar to that of the true nuts (Freitas & Naves, 2010; Venkatachalan & Sathe, 2006). Therefore, exotic edible seeds, either alone or with other nuts, are potential functional foods for healthy diets and food industry. Our study, in particular, contributes to the development of strategies for the conservation and sustainable use of these species as well as for the promotion of healthy diet, not only in Brazil but also in other countries where the nuts are appreciated (Jenab et al., 2006).

4. Conclusions

The baru almond, pequi almond, and cerrado cashew nut from the Brazilian Savanna, as well as the peanut, are foods with high nutritional (proteic and lipidic) content and energy value. In addition, these foods have an appreciable mineral content, mainly calcium, iron, and zinc, and baru and pequi almonds also have high contents of fibres. The baru almond and the cerrado cashew nut have a good amino acid profile and protein quality, and the protein value of baru almond is better than that of the peanut. On the other hand, the protein of the pequi almond is limited in lysine. We recommend the inclusion of these exotic foods in healthy diets and in food industry, and the baru almond and cerrado cashew nut as sources of complementary protein.

5. Acknowledgements

We acknowledge *Conselho Nacional de Desenvolvimento Científico e Tecnológico* (CNPq) and *Coordenação de Aperfeiçoamento de Pessoal de Nível Superior* (CAPES) for financial support and *Fundação de Amparo à Pesquisa do Estado de Goiás* (FAPEG) for scholarship support.

6. References

- Association of Official Analytical Chemists (AOAC). (1990). *Official methods of analysis*. (15th ed.). Arlington: AOAC.
- Bligh, E. G., & Dyer, W.J. (1959). A rapid method of total lipid extraction and purification. *Canadian Journal of Biochemistry and Physiology*, 37, 911–917.
- Correia, G. C., Naves, R. V., Rocha, M. R., Chaves, L. J., & Borges, J. D. (2008). Determinações físicas em frutos e sementes de baru (*Dipteryx alata* Vog., cajuzinho (*Anacardium othonianum* Rizz.) e pequi (*Caryocar brasiliense* Camb). *Bioscience Journal*, 24, 42–47.
- Fagbemi, T. N. (2008). The influence of processing techniques on the energy, ash properties and elemental composition of cashew nut (*Anacardium occidentale* Linn.). *Nutrition and Food Science*, 38, 136–145.
- Fernandes, D. C., Freitas, J. B., Czeder, L. P., & Naves, M. M. V. (2010). Nutritional composition and protein value of the baru (*Dipteryx alata* Vog.) almond from the Brazilian Savanna. *Journal of the Science of Food and Agriculture*, 90, 1650–1655.
- Food and Agriculture Organization (FAO) of the United Nations. Report of a Joint FAO/WHO Expert Consultation. (1991). *Protein quality evaluation* (FAO Food and Nutrition Paper n° 51). Rome: FAO.
- Food and Agriculture Organization (FAO) of the United Nations. (1970). *Amino-acid content of foods and biological data on proteins*. Rome: FAO.
- Freitas, J. B., & Naves, M. M. V. (2010). Composição química de nozes e sementes comestíveis e sua relação com a nutrição e saúde. *Revista de Nutrição*, 23, 269–279.
- Friedman, M. (1996). Nutritional value of proteins from different food sources. A review. *Journal of Agriculture and Food Chemistry*, 44, 6–29.
- Harper, A.E., Benevenga, N.J., Wohlhueter, R.M. (1970). Effects of ingestion of disproportionate amounts of amino acids. *Physiological Review*, 50, 428–458.
- Institute of Medicine (IOM). (2005). Food and Nutrition Board. Protein and amino acids. In IOM, *Dietary references intakes for energy, carbohydrate, fiber, fat, fatty acids, cholesterol, protein, and amino acids*. Washington, DC: The National Academies.
- Institute of Medicine (IOM). (2006). *Dietary reference intakes research synthesis: workshop summary*. Washington, DC: The National Academies.
- Jenab, M., Sabaté, J., Slimani, N., Ferrari, P., Mazuir, M., Casagrande, C., Dehaverng, G., Tjonneland, A., Olsen, A., Overvad, K., Boutron-Ruault, M. C., Clavel- Chapelon, F., Boeing, H., Weikert, C., Linseisen, J., Rohrmann, S., Trichopoulou, A., Naska, A., Palli, D., Sacerdote, C., Tumino, R., Mattiello, A., Pala, V., Bueno-de-Mesquita, H. B., Ocké, M. C., Peeters, P. H., Engeset, D., Skeie, G., Jakszyn, P., Ardanaz, E., Quirós, J. R., Chirilaque, M.

D., Martinez, C., Amiano, P., Berglund, G., Palmqvist, R., Van Guelpen, B., Bingham, S., Key, T. & Riboli, E. (2006). Consumption and portion sizes of tree nuts, peanuts and seeds in the European Prospective Investigation into Cancer and Nutrition (EPIC) cohorts from 10 European countries. *British Journal of Nutrition*, 96, S12–S23.

Judd, W. S., Campbell, C. S., Kellogg, E. A., Stevens, P. F., & Donoghue, M. J. (2002). Taxonomic evidence: structural and biochemical characters. In W. S. Judd, C. S. Campbell, E. A. Kellogg, P. F. Stevens, & M. J. Donoghue (Eds.), *Plant systematics: aphylogenetic approach* (pp. 55–104). Massachusetts: Sunderland.

Lima, A., Silva, A. M. O., Trindade, R. A., Torres, R. P., & Mancini-Filho, J. (2007). Composição química e compostos bioativos presentes na polpa e na amêndoa de pequi (*Caryocar brasiliense* Camb.). *Revista Brasileira de Fruticultura*, 29, 695–698.

López-Uriarte, P., Bulló, M., Casas-Agustench, P., Babio, N. & Salas-Salvado, J. (2009). Nuts and oxidation: a systematic review. *Nutrition Reviews*, 67, 497–508.

Lucas, B., & Sotelo, A. (1980). Effect of different alkalies, temperatures and hydrolysis times on tryptophan determination of pure proteins and foods. *Analytical Biochemistry*, 109, 192–197.

Melo Júnior, A. F., Carvalho, D., Póvoa, J. S. R., & Bearzoti, E. (2004). Estrutura genética de populações naturais de pequi (*Caryocar brasiliense* Camb.). *Scientia Forestalis*, 66, 56–65.

Merril, A. L., & Watt, B. K. (1973). *Energy value of foods: basis and derivation*. Washington, DC: United States Department of Agriculture.

Moore, S., Spackman, D. H., & Stein, W. H. (1958). Chromatography of amino acids on sulfonated polystyrene resins. *Analytical Chemistry*, 30, 1185–1190.

National Research Council (NRC). (1996). *Guide for the care and use of laboratory animals*. Washington, DC: National Academy Press.

Pellett, P. L., & Young, V. R. (1980). *Nutritional evaluation of protein foods*. Tokyo: The United Nations University.

Prosky, L., Asp, N., Schweizer, T. F., Devries, J. W., & Furda, I. (1988). Determination of insoluble, soluble and total dietary fiber in foods and foods products: interlaboratory study. *Journal of the Association of Official Analytical Chemists*, 71, 1017–1023.

Rabêlo, A. M. S., Torres, M. C. L., Geraldine, R. M., & Silveira, M. F. A. (2008). Extração, secagem e torrefação da amêndoa do pequi (*Caryocar brasiliense* Camb.). *Ciência e Tecnologia de Alimentos*, 28, 868–871.

Reeves, P. G., Nielsen, F. H., & Fahey JR., G. C. (1993). AIN-93 purified diets for laboratory rodents: final report of the American institute of Nutrition ad hoc writing committee on the reformulation of the AIN-76A rodent diet. *Journal of Nutrition*, 123, 1939–1951.

- Ruggeri, S., Cappelloni, M., Gambelli, L. L., & Carnovale, E. (1998). Chemical composition and nutritive value of nuts grown in Italy. *Italian Journal of Food Science*, 10, 243–252.
- Schaafsma, G. (2000). The protein digestibility-corrected amino acid score. *Journal of Nutrition*, 130, 1865S–1867S.
- Singh, B. & Singh, U. (1991). Peanut as a source of protein for human foods. *Plant Foods for Human Nutrition*, 41, 165–177.
- Takemoto, E., Okada, I. A., Garbelotti, M. L., Tavares, M., & Aued-Pimentel, S. (2001). Composição química da semente e do óleo de baru (*Dipteryx alata* Vog.) nativo do município de Pirenópolis, estado de Goiás. *Revista do Instituto Adolfo Lutz*, 60, 113–117.
- Togashi, M. & Sgarbieri, V. C. (1994). Caracterização química parcial do fruto do baru (*Dipteryx alata*, Vog.). *Ciência e Tecnologia de Alimentos*, 14, 85–95.
- Togashi, M., & Sgarbieri, V. C. (1995). Avaliação nutricional da proteína e do óleo de sementes de baru (*Dipteryx alata*, Vog.). *Ciência e Tecnologia de Alimentos*, 15, 66–69.
- Venkatachalam, M. & Sathe, S. K. (2006). Chemical composition of selected edible nut seeds. *Journal of Agricultural and Food Chemistry*, 54, 4705–4714.
- World Health Organization (WHO). Report of a Joint WHO/FAO/UNU Expert Consultation. (2007). *Protein and amino acid requirements in human nutrition*. (WHO Technical Report Series nº 935). Geneva: WHO.

Tables

Table 1

Ingredients and chemical composition of diets used in the biological assay

Component (g/100 g of diet)	Diet ^a						
	Reference	Control	Baru almond	Pequi almond	Cerrado cashew nut	Peanut	Protein- free
Casein	13.27	13.27	-	-	-	-	-
Baru almond	-	-	33.42	-	-	-	-
Pequi almond	-	-	-	33.73	-	-	-
Cerrado cashew nut	-	-	-	-	44.12	-	-
Peanut	-	-	-	-	-	33.79	-
L-Cystine	0.20	0.20	-	-	-	-	-
Soybean oil	6.58	14.58	-	-	-	-	7.00
Cellulose	5.00	5.00	0.52	4.26	3.37	2.30	5.00
Mineral mix	3.50	3.50	3.50	3.50	3.50	3.50	3.50
Vitamin mix	1.00	1.00	1.00	1.00	1.00	1.00	1.00
Choline bitartrate	0.25	0.25	0.25	0.25	0.25	0.25	0.25
Corn starch	70.20	62.20	61.31	57.26	47.76	59.16	83.25
Protein (g/100 g)	10.39	10.67	9.24	9.94	9.93	10.95	0.45
Lipids (g/100 g)	6.70	14.89	14.36	17.24	21.31	15.60	6.94
Energy value (kcal/100 g)	412.50	452.50	452.00	459.10	485.40	458.20	-

Reference: casein with 7% lipids; Control: casein with 15% lipids.

^aFeed formulation according to the AIN-93G diet (Reeves et al., 1993), with reduction of protein levels to 10% (w/w).

Table 2

Chemical composition and energy value of exotic almonds and nut compared to those of peanut

Component (g/100 g)*	Exotic almonds and nut			Peanut
	Baru almond	Pequi almond	Cerrado cashew nut	
Moisture	3.49 ± 0.08 ^{b,c}	4.97 ± 0.09 ^a	3.16 ± 0.11 ^c	3.53 ± 0.15 ^b
Proteins (N x 6.25)	29.92 ± 0.37 ^a	29.65 ± 0.55 ^a	22.67 ± 0.20 ^b	29.59 ± 0.05 ^a
Total lipids	41.95 ± 0.44 ^d	50.00 ± 0.66 ^a	47.79 ± 0.31 ^b	46.35 ± 0.26 ^c
Carbohydrates	12.25	0.40	19.86	13.06
Total dietary fibres	9.21 ± 0.21 ^b	10.44 ± 0.09 ^a	3.92 ± 0.05 ^d	5.20 ± 0.11 ^c
soluble fibres	2.03 ± 0.00 ^b	3.62 ± 0.13 ^a	1.17 ± 0.05 ^d	1.36 ± 0.02 ^c
insoluble fibres	7.18 ± 0.21 ^a	6.82 ± 0.21 ^b	2.76 ± 0.05 ^d	3.84 ± 0.09 ^c
Ash	3.18 ± 0.01 ^b	4.54 ± 0.02 ^a	2.60 ± 0.01 ^c	2.27 ± 0.01 ^d
Energy value (kcal/100 g)	546.23	570.20	600.23	587.75

^{a-d} Values marked with the same letter in the same row are not significantly different ($P > 0.05$ by Tukey's test).

*Data are mean ± standard deviation of 3 replicates, except carbohydrates, which were calculated by subtracting the values of the other components from 100.

Table 3

Mineral composition of exotic almonds and nut compared to that of peanut

Minerals* (mg/100 g)	Exotic almonds and nut			Peanut
	Baru almond	Pequi almond	Cerrado cashew nut	
Ca	110.94 ± 1.36 ^a	90.12 ± 0.71 ^b	64.05 ± 1.61 ^c	32.65 ± 1.20 ^d
Fe	3.57 ± 0.09 ^a	2.28 ± 0.13 ^b	3.89 ± 0.48 ^a	1.89 ± 0.24 ^b
K	980.35 ± 5.31 ^a	835.66 ± 15.46 ^b	556.16 ± 13.40 ^d	668.11 ± 3.73 ^c
Na	7.46 ± 1.51 ^b	5.68 ± 4.23 ^b	3.08 ± 1.14 ^b	19.47 ± 1.77 ^a
Mg	164.81 ± 1.29 ^c	452.11 ± 62.10 ^a	277.09 ± 2.90 ^b	190.47 ± 6.76 ^c
P	832.80 ± 2.66 ^c	2,214.46 ± 1.85 ^a	1,101.04 ± 10.87 ^b	856.29 ± 2.18 ^c
Se (µg/100 g)	0.37 ± 0.06 ^d	1.40 ± 0.01 ^b	1.02 ± 0.00 ^c	2.51 ± 0.00 ^a
Zn	4.29 ± 0.16 ^b	7.38 ± 0.78 ^a	4.98 ± 0.21 ^b	4.33 ± 1.03 ^b

^{a-d}Values marked by the same letter in the same row are not significantly different ($P > 0.05$ by Tukey's test).

*Data are mean ± standard deviation of 2 replicates.

Table 4

Amino acid composition and amino acid score (AAS), according to the WHO/FAO/UNU^a requirement pattern, of exotic almonds and nut compared to those of peanut

Amino acid (mg amino acid/g protein)	Requirement pattern*	Exotic almonds and nut ^b			Peanut
		Baru almond	Pequi almond	Cerrado cashew nut	
<u>Indispensable (essential)</u>					
His	16.0	23.4	28.4	23.3	26.8
Ile	31.0	32.5	27.7	41.9	32.7
Leu	61.0	74.4	64.5	73.1	67.1
Lys	48.0	66.4	30.3	44.6	43.8
Met+Cys	24.0	29.8	62.0	35.6	27.2
Phe+Tyr	41.0	88.5	67.1	86.7	102.3
Thr	25.0	55.3	21.7	36.0	31.8
Trp	6.6	11.2	10.0	16.5	7.1
Val	40.0	55.6	40.4	53.8	35.5
TOTAL	292.6	437.1	352.1	411.5	374.3
AAS (%)	100	105	63	93	89
<u>Dispensable (non-essential)</u>					
Asp	-	91.4	79.9	78.9	115.3
Glu	-	176.9	182.2	173.0	180.6
Ala	-	42.6	36.3	35.1	38.1
Arg	-	151.4	223.7	172.8	136.7
Gly	-	41.7	56.8	56.2	47.0
Pro	-	3.8	33.6	21.9	53.1
Ser	-	58.4	45.5	50.5	55.1
TOTAL		566.2	658.0	588.4	625.9

^aWHO (2007).

^bData are means of 2 replicates. Bold values indicate the first limiting amino acid.

Table 5

Food and protein intakes and protein value indexes of rats fed with different experimental diets

Parameter ^a	Diet*					
	Reference	Control	Baru almond	Pequi almond	Cerrado cashew nut	Peanut
Total food intake (g)	181.18 ± 15.32 ^a	168.57 ± 11.28 ^{a,b}	150.98 ± 10.95 ^b	97.02 ± 14.22 ^c	147.88 ± 4.85 ^b	154.08 ± 13.45 ^b
Total protein intake (g)	18.82 ± 1.74 ^a	17.99 ± 1.32 ^a	13.95 ± 1.01 ^c	9.64 ± 1.55 ^d	14.68 ± 0.53 ^{b,c}	16.87 ± 1.61 ^{a,b}
Weight gain (g)	63.53 ± 10.67 ^a	69.93 ± 4.60 ^a	40.87 ± 4.32 ^b	13.10 ± 4.56 ^c	38.38 ± 4.35 ^b	41.00 ± 5.70 ^b
NPR	3.81 ± 0.49 ^{a,b}	4.36 ± 0.41 ^a	3.53 ± 0.35 ^{b,c}	2.19 ± 0.29 ^e	3.17 ± 0.23 ^{c,d}	2.91 ± 0.20 ^d
RNPR (%)	100 ^a	100 ^a	86.44 ± 8.62 ^b	53.66 ± 7.09 ^d	77.63 ± 5.70 ^{b,c}	71.70 ± 4.87 ^c
True digestibility (%)	95.48 ± 2.25 ^{a,b}	96.85 ± 0.68 ^a	86.38 ± 0.75 ^c	88.11 ± 1.82 ^c	88.07 ± 1.21 ^c	92.74 ± 1.10 ^b
PDCAAS (%)	-	-	90.70 ± 0.79 ^a	55.51 ± 1.15 ^c	81.91 ± 1.13 ^b	82.54 ± 0.98 ^b

^{a-d}Values marked with the same letter in the same row are not significantly different ($P > 0.05$ by Tukey's test).

*Reference diet: casein with 7% lipids; Control diet: casein with 15% lipids. Data are mean ± standard deviation of six animals.

^aNPR: Net Protein Ratio (values were calculated considering the weight loss of rats fed with the protein-free diet: 8.20 ± 0.77 g); RNPR: Relative NPR; PDCAAS: Protein Digestibility-Corrected Amino Acid Score.

Figures

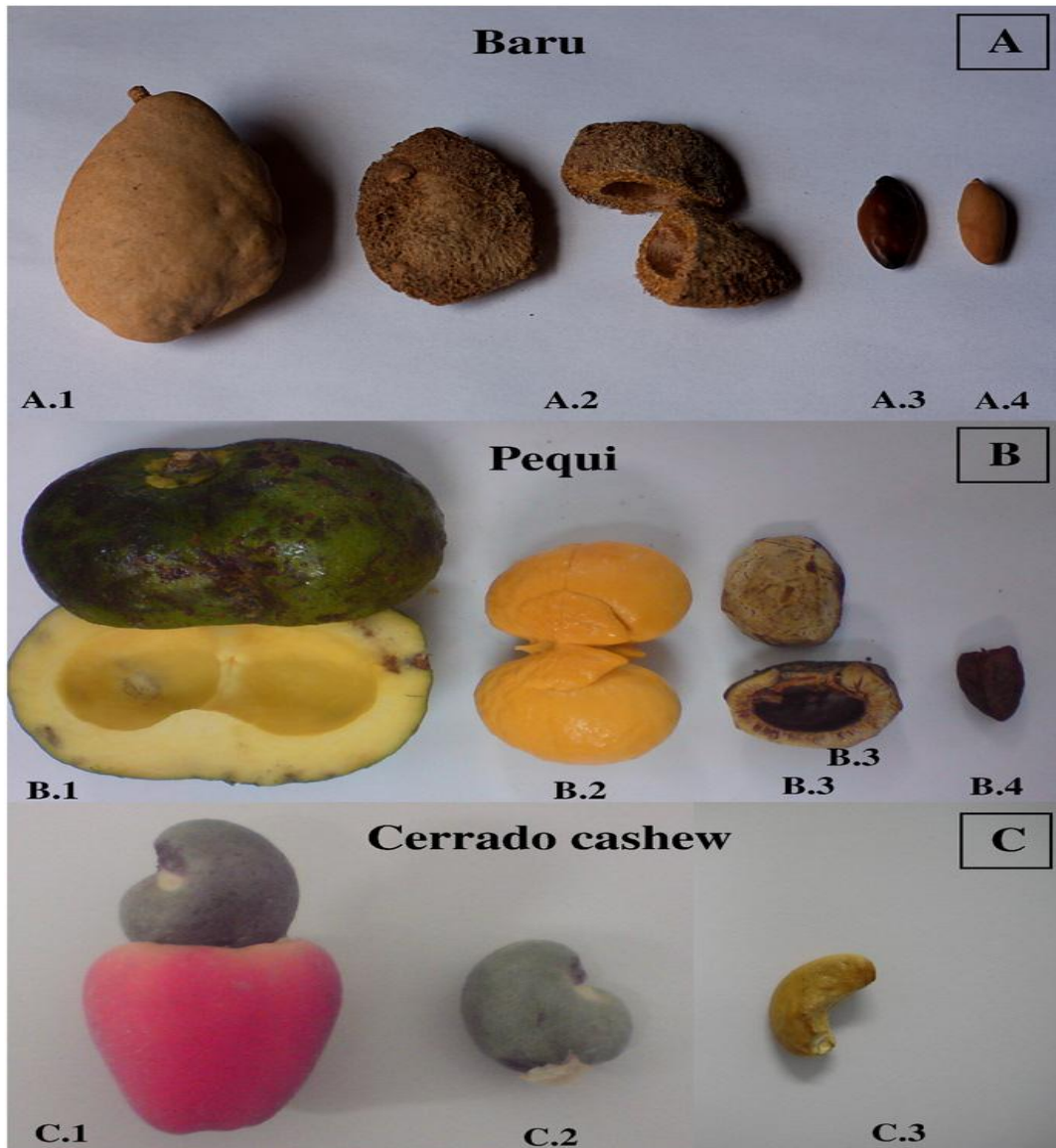


Fig. 1. Exotic almonds and nut from the Brazilian Savanna (fruits and almonds/nut).
 A: Baru fruit; A.1: Whole barufruit (ripe); A.2: Woody endocarp; A.3: Baruedible seed, also called baru almond; A.4: baru edible seed without the dark brown peel (usual form of consumption). B: Pequi fruit; B.1: Pequi greenish-brown exocarpus and external mesocarpus (white pulp); B.2: Pequi internal mesocarpus (yellow pulp); B.3: Pequi thorny endocarpus (without the internal mesocarpus) containing the edible seed; B.4: Pequi edible seed, also called pequi almond. C: Cerrado cashew fruit; C.1: True fruit and ripe pseudo-fruit (fleshy peduncle); C.2: Cashew true fruit containing the cashew nut; C.3: Cerrado cashew nut.

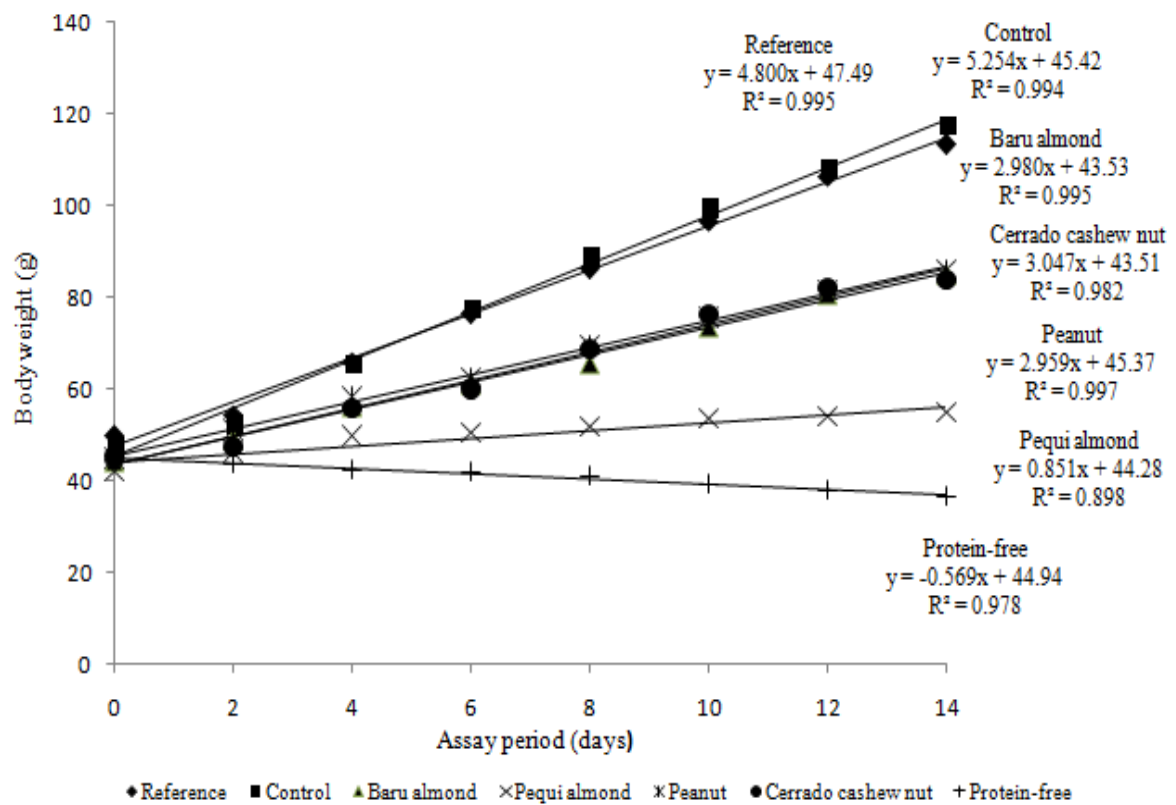


Fig. 2. Body weight of rats fed with different experimental diets for 14 days.

CAPÍTULO 3

CONSIDERAÇÕES FINAIS

As amêndoas de baru, de pequi e a castanha-de-caju-do-cerrado, bem como o amendoim, são alimentos de alta densidade energética e de nutrientes, especialmente proteínas, lipídios e minerais. A amêndoa de baru possui maior conteúdo total de aminoácidos essenciais em relação às demais sementes comestíveis e a noz, e ainda, possui maior qualidade protéica. Porém, a castanha-de-caju-do-cerrado e o amendoim também podem ser consideradas fontes de proteína de boa qualidade.

A inclusão destes alimentos nativos deve ser recomendada em dietas saudáveis para a população local e sugerimos que seja investigada a viabilidade da utilização destes alimentos na formulação de produtos industrializados, contribuindo para um melhor aproveitamento de alimentos regionais e para o uso racional das espécies.

ANEXOS

ANEXO A – Parecer do Comitê de Ética

SERVIÇO PÚBLICO FEDERAL
UNIVERSIDADE FEDERAL DE GOIÁS
PRÓ-REITORIA DE PESQUISA E PÓS-GRADUAÇÃO
COMITÊ DE ÉTICA EM PESQUISA



PROTOCOLO
153/2008

Goiânia, 14 de setembro de 2009

PARECER CONSUBSTANCIADO**I. IDENTIFICAÇÃO:**

Título do projeto: “Qualidade nutricional de frutos do cerrado e produtos derivados”

Pesquisador Responsável: Maria Margareth Veloso Naves

Pesquisadores Participantes: Mara Reis Silva/FANUT
Maria Sebastiana Silva/FEF

Local de realização: Faculdade de Nutrição/UFG

Área Temática: III

Informamos que o Comitê de Ética em Pesquisa da Universidade Federal de Goiás, após análise das adequações solicitadas, **Aprovou**, o projeto acima referido, e o mesmo foi considerado em acordo com os princípios éticos vigentes.

O pesquisador responsável deverá encaminhar ao CEP/UFG, relatórios da pesquisa, encerramento, conclusão (ões) e publicação (ões) de acordo com as recomendações da Resolução 196/96.


Profa. Dra. Rita Goreti Amaral
Coordenadora do CEP/UFG

ANEXO B – Comprovante atualizado Qualis Capes

WebQualis - Consulta Periódicos

qualis.capes.gov.br/webqualis/ConsultaPeriodicos.faces

WebQualis

Ajuda?

CONSULTAR DOCUMENTOS DE ÁREA E-MAIL DOS COORDENADORES LISTA COMPLETA LOGIN

Selecione o tipo de detalhamento da pesquisa:

Por ISSN do Periódico Por Título do Periódico Por Classificação / Área de Avaliação

food research international CONSULTAR

* Ano-Base 2008

Título: food research international

ISSN	Título	Estrato	Área de Avaliação
0963-9969	Food Research International	A1	ENGENHARIAS III
0963-9969	Food Research International	A2	CIÊNCIA DE ALIMENTOS
0963-9969	Food Research International	A2	ENGENHARIAS II
0963-9969	Food Research International	B1	CIÊNCIAS BIOLÓGICAS I
0963-9969	Food Research International	B1	ENGENHARIAS IV
0963-9969	Food Research International	B1	FARMÁCIA
0963-9969	Food Research International	B1	MEDICINA II
0963-9969	Food Research International	B1	QUÍMICA
0963-9969	Food Research International	B2	BIOTECNOLOGIA
0963-9969	Food Research International	B2	CIÊNCIAS BIOLÓGICAS III

Página: 1 de 1

PT 23:34

ANEXO C – Normas de publicação do periódico *Food Research International*

Ethics in Publishing

For information on Ethics in Publishing and Ethical guidelines for journal publication see ➡ <http://www.elsevier.com/publishingethics> and ➡ <http://www.elsevier.com/ethicalguidelines>.

Policy and ethics

The work described in your article must have been carried out in accordance with *The Code of Ethics of the World Medical Association (Declaration of Helsinki) for experiments involving humans* ➡ <http://www.wma.net/e/policy/b3.htm>; *EC Directive 86/609/EEC for animal experiments* ➡ http://ec.europa.eu/environment/chemicals/lab_animals/legislation_en.htm; *Uniform Requirements for manuscripts submitted to Biomedical journals* ➡ <http://www.icmje.org>. This must be stated at an appropriate point in the article.

Conflict of interest

All authors are requested to disclose any actual or potential conflict of interest including any financial, personal or other relationships with other people or organizations within three years of beginning the submitted work that could inappropriately influence, or be perceived to influence, their work. See also ➡ <http://www.elsevier.com/conflictsofinterest>.

Submission declaration

Submission of an article implies that the work described has not been published previously (except in the form of an abstract or as part of a published lecture or academic thesis), that it is not under consideration for publication elsewhere, that its publication is approved by all authors and tacitly or explicitly by the responsible authorities where the work was carried out, and that, if accepted, it will not be published elsewhere including electronically in the same form, in English or in any other language, without the written consent of the copyright-holder.

Contributors

Each author is required to declare his or her individual contribution to the article: all authors must have materially participated in the research and/or article preparation, so roles for all authors should be described. The statement that all authors have approved the final article should be true and included in the disclosure.

Copyright

Upon acceptance of an article, authors will be asked to complete a 'Journal Publishing Agreement' (for more information on this and copyright see ➡ <http://www.elsevier.com/copyright>). Acceptance of the agreement will ensure the widest possible dissemination of information. An e-mail will be sent to the corresponding author confirming receipt of the manuscript together with a 'Journal Publishing Agreement' form or a link to the online version of this agreement.

Subscribers may reproduce tables of contents or prepare lists of articles including abstracts for internal circulation within their institutions. Permission of the Publisher is required for resale or distribution outside the institution and for all other derivative works, including compilations and translations (please consult ➡ <http://www.elsevier.com/permissions>). If excerpts from other copyrighted works are included, the author(s) must obtain written permission from the copyright owners and credit the source(s) in the article. Elsevier has preprinted forms for use by authors in these cases: please consult ➡ <http://www.elsevier.com/permissions>.

Retained author rights

As an author you (or your employer or institution) retain certain rights; for details you are referred to: ➡ <http://www.elsevier.com/authorsrights>.

Role of the funding source

You are requested to identify who provided financial support for the conduct of the research and/or preparation of the article and to briefly describe the role of the sponsor(s), if any, in study design; in the collection, analysis and interpretation of data; in the writing of the report; and in the decision to submit the paper for publication. If the funding source(s) had no such involvement then this should be stated. Please see <http://www.elsevier.com/funding>.

Funding body agreements and policies

Elsevier has established agreements and developed policies to allow authors whose articles appear in journals published by Elsevier, to comply with potential manuscript archiving requirements as specified as conditions of their grant awards. To learn more about existing agreements and policies please visit <http://www.elsevier.com/fundingbodies>.

Sponsored articles

This journal offers you the choice of making your article freely available to all on Elsevier's electronic publishing platforms. The charge for article sponsorship is \$3,000, which is necessary to offset publishing costs. To prevent any conflict of interest, you can only make this choice after receiving notification that your article has been accepted for publication. Full details of the sponsored Open Access options available to you and your funding body can be found here: <http://www.elsevier.com/locate/sponsoredarticles>. Whatever access option you choose, you retain many rights as an author, including the right to post a revised personal version of your article on your own website. More information can be found here: <http://www.elsevier.com/authorsrights>.

Language and language services

Please write your text in good English (American or British usage is accepted, but not a mixture of these). Authors who require information about language editing and copyediting services pre- and post-submission please visit <http://www.elsevier.com/languageediting> or our customer support site at <http://support.elsevier.com> for more information.

Submission

Submission to this journal proceeds totally online and you will be guided stepwise through the creation and uploading of your files. The system automatically converts source files to a single PDF file of the article, which is used in the peer-review process. Please note that even though manuscript source files are converted to PDF files at submission for the review process, these source files are needed for further processing after acceptance. All correspondence, including notification of the Editor's decision and requests for revision, takes place by e-mail removing the need for a paper trail.

Referees

Please submit, with the manuscript, the names, addresses and e-mail addresses of 3 potential referees. Note that the editor retains the sole right to decide whether or not the suggested reviewers are used.



Preparation

Use of wordprocessing software

It is important that the file be saved in the native format of the wordprocessor used. The text should be in single-column format. Keep the layout of the text as simple as possible. Most formatting codes will be removed and replaced on processing the article. In particular, do not use the wordprocessor's options to justify text or to hyphenate words. However, do use bold face, italics, subscripts, superscripts etc. Do not embed "graphically designed" equations or tables, but prepare these using the wordprocessor's facility. When preparing tables, if you are using a table grid, use only one grid for

each individual table and not a grid for each row. If no grid is used, use tabs, not spaces, to align columns. The electronic text should be prepared in a way very similar to that of conventional manuscripts (see also the Guide to Publishing with Elsevier: <http://www.elsevier.com/guidepublication>). Do not import the figures into the text file but, instead, indicate their approximate locations directly in the electronic text and on the manuscript. See also the section on Electronic illustrations.

To avoid unnecessary errors you are strongly advised to use the "spell-check" and "grammar-check" functions of your wordprocessor.

All pages of the manuscript must be numbered. All lines must be numbered continuously throughout the manuscript.

Article structure

Subdivision - numbered sections

Divide your article into clearly defined and numbered sections. Subsections should be numbered 1.1 (then 1.1.1, 1.1.2, ...), 1.2, etc. (the abstract is not included in section numbering). Use this numbering also for internal cross-referencing: do not just refer to "the text". Any subsection may be given a brief heading. Each heading should appear on its own separate line.

Introduction

State the objectives of the work and provide an adequate background, avoiding a detailed literature survey or a summary of the results.

Material and methods

Provide sufficient detail to allow the work to be reproduced. Methods already published should be indicated by a reference: only relevant modifications should be described.

Experimental

Provide sufficient detail to allow the work to be reproduced. Methods already published should be indicated by a reference: only relevant modifications should be described.

Theory/calculation

A Theory section should extend, not repeat, the background to the article already dealt with in the Introduction and lay the foundation for further work. In contrast, a Calculation section represents a practical development from a theoretical basis.

Results

Results should be clear and concise.

Discussion

This should explore the significance of the results of the work, not repeat them. A combined Results and Discussion section is often appropriate. Avoid extensive citations and discussion of published literature.

Conclusions

The main conclusions of the study may be presented in a short Conclusions section, which may stand alone or form a subsection of a Discussion or Results and Discussion section.

Appendices

If there is more than one appendix, they should be identified as A, B, etc. Formulae and equations in appendices should be given separate numbering: Eq. (A.1), Eq. (A.2), etc.; in a subsequent appendix, Eq. (B.1) and so on. Similarly for tables and figures: Table A.1; Fig. A.1, etc.

Essential title page information

- **Title.** Concise and informative. Titles are often used in information-retrieval systems. Avoid abbreviations and formulae where possible.
- **Author names and affiliations.** Where the family name may be ambiguous (e.g., a double name), please indicate this clearly. Present the authors' affiliation addresses (where the actual work was

done) below the names. Indicate all affiliations with a lower-case superscript letter immediately after the author's name and in front of the appropriate address. Provide the full postal address of each affiliation, including the country name, and, if available, the e-mail address of each author.

• **Corresponding author.** Clearly indicate who will handle correspondence at all stages of refereeing and publication, also post-publication. **Ensure that telephone and fax numbers (with country and area code) are provided in addition to the e-mail address and the complete postal address.**

• **Present/permanent address.** If an author has moved since the work described in the article was done, or was visiting at the time, a "Present address" (or "Permanent address") may be indicated as a footnote to that author's name. The address at which the author actually did the work must be retained as the main, affiliation address. Superscript Arabic numerals are used for such footnotes.

Abstract

A concise and factual abstract is required. The abstract should state briefly the purpose of the research, the principal results and major conclusions. An abstract is often presented separately from the article, so it must be able to stand alone. For this reason, References should be avoided, but if essential, then cite the author(s) and year(s). Also, non-standard or uncommon abbreviations should be avoided, but if essential they must be defined at their first mention in the abstract itself.

Keywords

Immediately after the abstract, provide a maximum of 6 keywords, using American spelling and avoiding general and plural terms and multiple concepts (avoid, for example, "and", "of"). Be sparing with abbreviations: only abbreviations firmly established in the field may be eligible. These keywords will be used for indexing purposes.

Abbreviations

Define abbreviations that are not standard in this field in a footnote to be placed on the first page of the article. Such abbreviations that are unavoidable in the abstract must be defined at their first mention there, as well as in the footnote. Ensure consistency of abbreviations throughout the article.

Acknowledgements

Collate acknowledgements in a separate section at the end of the article before the references and do not, therefore, include them on the title page, as a footnote to the title or otherwise. List here those individuals who provided help during the research (e.g., providing language help, writing assistance or proof reading the article, etc.).

Units

Follow internationally accepted rules and conventions: use the international system of units (SI). If other units are mentioned, please give their equivalent in SI.

Math formulae

Present simple formulae in the line of normal text where possible and use the solidus (/) instead of a horizontal line for small fractional terms, e.g., X/Y. In principle, variables are to be presented in italics. Powers of e are often more conveniently denoted by exp. Number consecutively any equations that have to be displayed separately from the text (if referred to explicitly in the text).

Footnotes

Footnotes should be used sparingly. Number them consecutively throughout the article, using superscript Arabic numerals. Many wordprocessors build footnotes into the text, and this feature may be used. Should this not be the case, indicate the position of footnotes in the text and present the footnotes themselves separately at the end of the article. Do not include footnotes in the Reference list.

Table footnotes

Indicate each footnote in a table with a superscript lowercase letter.

Artwork

Image manipulation

Whilst it is accepted that authors sometimes need to manipulate images for clarity, manipulation for purposes of deception or fraud will be seen as scientific ethical abuse and will be dealt with accordingly. For graphical images, this journal is applying the following policy: no specific feature within an image may be enhanced, obscured, moved, removed, or introduced. Adjustments of brightness, contrast, or color balance are acceptable if and as long as they do not obscure or eliminate any information present in the original. Nonlinear adjustments (e.g. changes to gamma settings) must be disclosed in the figure legend.

Electronic artwork

General points

- Make sure you use uniform lettering and sizing of your original artwork.
- Save text in illustrations as "graphics" or enclose the font.
- Only use the following fonts in your illustrations: Arial, Courier, Times, Symbol.
- Number the illustrations according to their sequence in the text.
- Use a logical naming convention for your artwork files.
- Provide captions to illustrations separately.
- Produce images near to the desired size of the printed version.
- Submit each figure as a separate file.

A detailed guide on electronic artwork is available on our website:

➡ <http://www.elsevier.com/artworkinstructions>

You are urged to visit this site; some excerpts from the detailed information are given here.

Formats

Regardless of the application used, when your electronic artwork is finalised, please "save as" or convert the images to one of the following formats (note the resolution requirements for line drawings, halftones, and line/halftone combinations given below):

EPS: Vector drawings. Embed the font or save the text as "graphics".

TIFF: color or grayscale photographs (halftones): always use a minimum of 300 dpi.

TIFF: Bitmapped line drawings: use a minimum of 1000 dpi.

TIFF: Combinations bitmapped line/half-tone (color or grayscale): a minimum of 500 dpi is required.

DOC, XLS or PPT: If your electronic artwork is created in any of these Microsoft Office applications please supply "as is".

Please do not:

- Supply embedded graphics in your wordprocessor (spreadsheet, presentation) document;
- Supply files that are optimised for screen use (like GIF, BMP, PICT, WPG); the resolution is too low;
- Supply files that are too low in resolution;
- Submit graphics that are disproportionately large for the content.

Color artwork

Please make sure that artwork files are in an acceptable format (TIFF, EPS or MS Office files) and with the correct resolution. If, together with your accepted article, you submit usable color figures then Elsevier will ensure, at no additional charge, that these figures will appear in color on the Web (e.g., ScienceDirect and other sites) regardless of whether or not these illustrations are reproduced in color in the printed version. **For color reproduction in print, you will receive information regarding the costs from Elsevier after receipt of your accepted article.** Please indicate your preference for color in print or on the Web only. For further information on the preparation of electronic artwork, please see ➡ <http://www.elsevier.com/artworkinstructions>.

Please note: Because of technical complications which can arise by converting color figures to "gray scale" (for the printed version should you not opt for color in print) please submit in addition usable black and white versions of all the color illustrations.

Figure captions

Ensure that each illustration has a caption. Supply captions separately, not attached to the figure. A caption should comprise a brief title (**not** on the figure itself) and a description of the illustration. Keep text in the illustrations themselves to a minimum but explain all symbols and abbreviations used.

Tables

Number tables consecutively in accordance with their appearance in the text. Place footnotes to tables below the table body and indicate them with superscript lowercase letters. Avoid vertical rules. Be sparing in the use of tables and ensure that the data presented in tables do not duplicate results described elsewhere in the article.

References

Citation in text

Please ensure that every reference cited in the text is also present in the reference list (and vice versa). Any references cited in the abstract must be given in full. Unpublished results and personal communications are not recommended in the reference list, but may be mentioned in the text. If these references are included in the reference list they should follow the standard reference style of the journal and should include a substitution of the publication date with either "Unpublished results" or "Personal communication". Citation of a reference as "in press" implies that the item has been accepted for publication.

Web references

As a minimum, the full URL should be given and the date when the reference was last accessed. Any further information, if known (DOI, author names, dates, reference to a source publication, etc.), should also be given. Web references can be listed separately (e.g., after the reference list) under a different heading if desired, or can be included in the reference list.

References in a special issue

Please ensure that the words 'this issue' are added to any references in the list (and any citations in the text) to other articles in the same Special Issue.

Reference management software

This journal has standard templates available in key reference management packages EndNote (<http://www.endnote.com/support/enstyles.asp>) and Reference Manager (<http://refman.com/support/rmstyles.asp>). Using plug-ins to wordprocessing packages, authors only need to select the appropriate journal template when preparing their article and the list of references and citations to these will be formatted according to the journal style which is described below.

Reference style

Text: Citations in the text should follow the referencing style used by the American Psychological Association. You are referred to the Publication Manual of the American Psychological Association, Sixth Edition, ISBN 978-1-4338-0561-5, copies of which may be ordered from <http://books.apa.org/books.cfm?id=4200067> or APA Order Dept., P.O.B. 2710, Hyattsville, MD 20784, USA or APA, 3 Henrietta Street, London, WC3E 8LU, UK. Details concerning this referencing style can also be found at <http://linguistics.byu.edu/faculty/henrichsen/apa/apa01.html>.

List: references should be arranged first alphabetically and then further sorted chronologically if necessary. More than one reference from the same author(s) in the same year must be identified by the letters "a", "b", "c", etc., placed after the year of publication.

Examples:

Reference to a journal publication:

Van der Geer, J., Hanraads, J. A. J., & Lupton, R. A. (2000). The art of writing a scientific article. *Journal of Scientific Communications*, 163, 51–59.

Reference to a book:

Strunk, W., Jr., & White, E. B. (1979). *The elements of style*. (3rd ed.). New York: Macmillan, (Chapter 4).

Reference to a chapter in an edited book:

Mettam, G. R., & Adams, L. B. (1994). How to prepare an electronic version of your article. In B. S. Jones, & R. Z. Smith (Eds.), *Introduction to the electronic age* (pp. 281–304). New York: E-Publishing Inc.

Video data

Elsevier accepts video material and animation sequences to support and enhance your scientific

research. Authors who have video or animation files that they wish to submit with their article are strongly encouraged to include these within the body of the article. This can be done in the same way as a figure or table by referring to the video or animation content and noting in the body text where it should be placed. All submitted files should be properly labeled so that they directly relate to the video file's content. In order to ensure that your video or animation material is directly usable, please provide the files in one of our recommended file formats with a maximum size of 10 MB. Video and animation files supplied will be published online in the electronic version of your article in Elsevier Web products, including ScienceDirect: ➡ <http://www.sciencedirect.com>. Please supply 'stills' with your files: you can choose any frame from the video or animation or make a separate image. These will be used instead of standard icons and will personalize the link to your video data. For more detailed instructions please visit our video instruction pages at ➡ <http://www.elsevier.com/artworkinstructions>. Note: since video and animation cannot be embedded in the print version of the journal, please provide text for both the electronic and the print version for the portions of the article that refer to this content.

Supplementary data

Elsevier accepts electronic supplementary material to support and enhance your scientific research. Supplementary files offer the author additional possibilities to publish supporting applications, high-resolution images, background datasets, sound clips and more. Supplementary files supplied will be published online alongside the electronic version of your article in Elsevier Web products, including ScienceDirect: ➡ <http://www.sciencedirect.com>. In order to ensure that your submitted material is directly usable, please provide the data in one of our recommended file formats. Authors should submit the material in electronic format together with the article and supply a concise and descriptive caption for each file. For more detailed instructions please visit our artwork instruction pages at ➡ <http://www.elsevier.com/artworkinstructions>.

Submission checklist

It is hoped that this list will be useful during the final checking of an article prior to sending it to the journal's Editor for review. Please consult this Guide for Authors for further details of any item.

Ensure that the following items are present:

One Author designated as corresponding Author:

- E-mail address
- Full postal address
- Telephone and fax numbers

All necessary files have been uploaded

- Keywords
- All figure captions
- All tables (including title, description, footnotes)

Further considerations

- Manuscript has been "spellchecked" and "grammar-checked"
- References are in the correct format for this journal
- All references mentioned in the Reference list are cited in the text, and vice versa
- Permission has been obtained for use of copyrighted material from other sources (including the Web)
- Color figures are clearly marked as being intended for color reproduction on the Web (free of charge) and in print or to be reproduced in color on the Web (free of charge) and in black-and-white in print
- If only color on the Web is required, black and white versions of the figures are also supplied for printing purposes

For any further information please visit our customer support site at <http://support.elsevier.com>.